

Package ‘tcpl’

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Title ToxCast Data Analysis Pipeline

Version 3.0.1

Description A set of tools for processing and modeling high-throughput and high-content chemical screening data. The package was developed for the the chemical screening data generated by the US EPA ToxCast program, but can be used for diverse chemical screening efforts.

URL <https://github.com/USEPA/CompTox-ToxCast-tcpl>

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`.buildAssayQ` *Generate query for assay information*

Description

`.buildAssayQ` generates a query string to load assay information

Usage

```
.buildAssayQ(out, tblo, fld = NULL, val = NULL, add.fld = NULL)
```

Arguments

<code>out</code>	Character, the default fields to include
<code>tblo</code>	Integer, the order to send the fields to prepOutput
<code>fld</code>	Character, the field(s) to query/subset on
<code>val</code>	List, vectors of values for each field to query/subset on. Must be in the same order as 'fld'.
<code>add.fld</code>	Character, additional field(s) to include, but not query/ subset on

Value

A character containing the query to send to tcplQuery

`.convertNames` *Convert assay names to their abbreviations*

Description

`.convertNames` converts the assay names as they appear in the tcpl database to their respective abbreviations

Usage

```
.convertNames(names)
```

Arguments

<code>names</code>	Character, strings to convert
--------------------	-------------------------------

Value

The same character vector given with any name strings converted to the abbreviated version

.load6DR *Load data for tcpl6*

Description

.load6DR loads dose-response data for tcpl6.

Usage

.load6DR(ae)

Arguments

ae String acid to query on

.plateHeat *Plot plate heatmap*

Description

Plot plate heatmap, to be used with tcplPlotPlate

Usage

.plateHeat(vals, rowi, coli, wllt, wllq, rown, coln, main, arng)

Arguments

vals Numeric, the well values
rowi Integer, the row index
coli Integer, the column index
wllt Character, the well type
wllq Logical, the well quality
rown Integer, the number of rows on the plate
coln Integer, the number of columns on the plate
main Character of length 1, the title/main
arng Numeric of length 2, the minimum and maximum values to constrain the color scale

Note

Optimized for an output with height = 20/3, width = 10, and pointsize = 10

`.prepField` *Paste appropriate table name to field name*

Description

Paste appropriate table name to field name

Usage

```
.prepField(fld, tbl, db)
```

Arguments

<code>fld</code>	Character, the table fields
<code>tbl</code>	Character, the possible tables
<code>db</code>	Character, the database containing the tables

Details

The function loops through the given tables, and for each field *i* it assigns the last table containing *i* to *i*. ORDER OF FLD MATTERS!!

`blineShift` *Shift the baseline to 0*

Description

`blineShift` Takes in dose-response data and shifts the baseline to 0 based on the window.

Usage

```
blineShift(resp, logc, wndw)
```

Arguments

<code>resp</code>	Numeric, the response values
<code>logc</code>	Numeric, the log10 concentration values
<code>wndw</code>	Numeric, the threshold window

Value

A numeric vector containing the shifted response values

Note

This function is not exported and is not intended to be used by the user.

See Also[mc3_mthds](#), [mc3](#)

chdat	<i>Chemical library of tested chemicals in the example datasets with the corresponding sample IDs.</i>
-------	--

Description

Chemical library of tested chemicals in the example datasets with the corresponding sample IDs.

Usage

```
chdat
```

Format

A data frame with 6 rows and 6 variables:

spid sample ID
casn Chemical Abstract Service(CAS) number
chnm chemical name
dsstox_substance_id chemical-specific DTXSID
code CAS number compressed into numeric string
chid unique chemical ID number for tcpl

Source

ToxCast database

check_tcpl_db_schema	<i>Function that checks if the most recent v3 table schema is used in the database schema</i>
----------------------	---

Description

Function that checks if the most recent v3 table schema is used in the database schema

Usage

```
check_tcpl_db_schema()
```

Value

boolean TRUE if param tables are listed in schema FALSE otherwise

Examples

```
## Not run:
#connect to database first with tcplConf
tcplConf(user=user,
  pass= pass,
  db=dbname,
  drvr='MySQL',
  host=hostname)

#check if it is part of the new schema
new_schema <- check_tcpl_db_schema()

## End(Not run)
```

Configure functions *Functions for configuring the tcpl package*

Description

These functions are used to configure the tcpl settings.

Usage

```
tcplConf(drvr = NULL, user = NULL, pass = NULL, host = NULL, db = NULL, ...)

tcplConfDefault()

tcplConfExample()

tcplConfList(show.pass = FALSE)

tcplConfLoad(list.new = TRUE)

tcplConfReset()

tcplConfSave()
```

Arguments

<code>drvr</code>	Character of length 1, which database driver to use
<code>user</code>	Character of length 1, the database server username
<code>pass</code>	Character of length 1, the database server password
<code>host</code>	Character of length 1, the database server
<code>db</code>	Character of length 1, the name of the tcpl database
<code>...</code>	Additional arguments that should be passed to dbConnect function
<code>show.pass</code>	Logical, should the password be returned
<code>list.new</code>	Logical of length 1, should the new settings be printed?

Details

Currently, the tcpl package only supports the "MySQL" and "tcplLite" database drivers.

The settings can be stored in a configuration file to make the using the package more user-friendly. To create the configuration file, the user must first create a system environment variable ('TCPL_CONF') that points to the file. There is more information about system environment variables in [Startup](#) and [Sys.getenv](#). Briefly, the user needs to modify the '.Renviro' file in their home directory. If the file does not exist, create it, and add the following line:

```
TCPL_CONF=path/to/confFile.conf
```

Here 'path/to/confFile.conf' can be any path to a file. One suggestion would be to include .tcplConf in the home directory, e.g. TCPL_CONF=~/.tcplConf. Note, '~' may not indicate the home directory on every operating system. Once the environment variable is added, the user can change the settings using tcplConf, then save the settings to the file given by the TCPL_CONF environment variable running tcplConfSave().

tcplConf changes options to set the tcpl-specific options, most importantly to configure the connection to the tcpl databases. tcplConf will only change non-null values, and can be used to change a single value if needed.

tcplConfSave modifies the configuration file to reflect the current tcpl settings.

tcplConfList lists the values assigned to the tcpl global options.

tcplConfLoad updates the tcpl settings to reflect the current configuration file.

tcplConfDefault changes the options to reflect the default settings for the example tcplLite database, i.e. local directory, but does not alter the configuration file.

tcplConfReset is used to generate the initial configuration script, and can be used to reset or regenerate the configuration script by the user.

flareFunc

Calculate the weighted mean of a square to detect plate flares

Description

flareFunc calculates the weighted mean of square regions to detect plate flares.

Usage

```
flareFunc(val, coli, rowi, apid, r)
```

Arguments

val	Numeric, the well values
coli	Integer, the well column index
rowi	Integer, the well row index
apid	Character, the assay plate id
r	Integer, the number of wells from the center well (in one direction) to make the square

See Also

[MC6_Methods](#), [Method functions](#), [mc6](#)

Hill model utilites *Functions to solve the Hill model*

Description

These functions solve for Hill model parameters.

Usage

```
tcplHillACXX(XX, tp, ga, gw, bt = 0)
```

```
tcplHillConc(val, tp, ga, gw, bt = 0)
```

```
tcplHillVal(logc, tp, ga, gw, bt = 0)
```

Arguments

XX	Numeric, the activity level (percentage of the top value)
tp	Numeric, the top value from the Hill model
ga	Numeric, the logAC50 value from the Hill model
gw	Numeric, the Hill coefficient from the Hill model
bt	Numeric, the bottom value from the Hill model
val	Numeric, the activity value
logc	Numeric, the log concentration

Details

tcplHillVal computes the value of the Hill model for a given log concentration.

tcplHillACXX computes the activity concentration for a Hill model for a given activity level.

tcplHillConc computes the Hill model concentration for a given value.

Examples

```
## The following code gives examples for a Hill model with a top of 50,
## bottom of 0, AC50 of 1 and Hill coefficient of 1.
## tcplHillVal calculates activity value given a concentration. tcplHillVal
## will return the tp/2 when logc equals ga:
tcplHillVal(logc = 1, tp = 50, ga = 1, gw = 1, bt = 0)

## Here, tcplHillConc returns the concentration where the value equals 20
tcplHillConc(val = 20, tp = 50, ga = 1, gw = 1, bt = 0)

## Note how this differs from tcplHillACXX:
```

```

tcplHillACXX(XX = 20, tp = 50, ga = 1, gw = 1, bt = 0)

## tcplHillACXX is based on the top value and allows the user to calculate
## specific activity concentrations based on a percentage of the top value

## For example, we can calculate the value for the concentration 0.25, then
## use that value to check the other two functions.

value <- tcplHillVal(logc = 0.25, tp = 50, ga = 1, gw = 1, bt = 0)
c1 <- tcplHillConc(val = value, tp = 50, ga = 1, gw = 1, bt = 0)
c2 <- tcplHillACXX(XX = value/50*100, tp = 50, ga = 1, gw = 1, bt = 0)
all.equal(0.25, c1, c2)

## Notice, the value had to be transformed to a percentage of the top value
## when using tcplHillACXX

```

interlaceFunc	<i>Calculate the weighted mean of a square to detect interlace effect</i>
---------------	---

Description

interlaceFunc calculates the distance weighted mean of square regions from a 384-well plate that is interlaced onto a 1536 well plate to detect non-random signals coming from the source plate

Usage

```
interlaceFunc(val, intq, coli, rowi, apid, r)
```

Arguments

val	Numeric, the well values
intq	Numeric, interlace quadrant
coli	Integer, the well column index
rowi	Integer, the well row index
apid	Character, the assay plate id
r	Integer, the number of wells from the center well (in one direction) to make the square

See Also

[MC6_Methods](#), [Method functions](#), [mc6](#)

<code>is.odd</code>	<i>Check for odd numbers</i>
---------------------	------------------------------

Description

`is.odd` takes an integer vector, `x`, and returns TRUE for odd integers.

Usage

```
is.odd(x)
```

Arguments

`x` An integer

Value

TRUE for odd integers and FALSE for even integers.

See Also

Other tcpl abbreviations: [lu\(\)](#), [lw\(\)](#), [sink.reset\(\)](#)

Load assay information

Functions for loading assay information

Description

These functions query the tcpl databases and returns a `data.table` with assay ID and name information. More information about the assay hierarchy is available in the overview vignette.

Usage

```
tcplLoadAcid(fld = NULL, val = NULL, add.fld = NULL)
```

```
tcplLoadAeid(fld = NULL, val = NULL, add.fld = NULL)
```

```
tcplLoadAid(fld = NULL, val = NULL, add.fld = NULL)
```

```
tcplLoadAsid(fld = NULL, val = NULL, add.fld = NULL)
```

Arguments

<code>fld</code>	Character, the field(s) to query/subset on
<code>val</code>	List, vectors of values for each field to query/subset on. Must be in the same order as 'fld'.
<code>add.fld</code>	Character, additional field(s) to include, but not query/ subset on

Details

Each element in the assay hierarchy has its own function, loading the ID and name for the given assay element. For example, `tcplLoadAsid` will return the assay source ID (`asid`) and assay source name (`asnm`).

Value

A `data.table` containing the ID, name, and any additional fields.

Examples

```
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcplConfList()
TCPLlite <- file.path(system.file(package = "tcpl"), "example")
tcplConf(db = TCPLlite, user = NA, host = NA, drvr = "tcplLite")

## The load assay functions can be used without any parameters to list the
## full list of registered assay elements:
tcplLoadAsid()
tcplLoadAeid()

## Similarly, the user can add fields without doing any element selection:
tcplLoadAeid(add.fld = c("asid", "aid", "acid"))

## Or, the user can look only at a subset:
tcplLoadAeid(fld = "aeid", val = 1, add.fld = "asid")

## The field can be any value in one of the corresponding assay element
## tables, but the functions also recognize the abbreviated version of
## the name fields.
tcplListFlds("assay")
a1 <- tcplLoadAeid(fld = "anm", val = "Steroidogenesis")
a2 <- tcplLoadAeid(fld = "assay_name", val = "Steroidogenesis")
identical(a1, a2)

## Reset configuration
options(conf_store)
```

lu	<i>Abbreviation for</i> length(unique(x))
----	---

Description

lu takes a logical vector, x, and returns length(unique(x)).

lu takes a logical vector, x, and returns length(unique(x)).

Usage

```
lu(x)
```

```
lu(x)
```

Arguments

x A logical

Value

The unique of the TRUE values in x

The unique of the TRUE values in x

See Also

[unique, which](#)

[unique, which](#)

Other tcpl abbreviations: [is.odd\(\)](#), [lw\(\)](#), [sink.reset\(\)](#)

Other tcpl abbreviations: [is.odd\(\)](#), [lw\(\)](#), [sink.reset\(\)](#)

lw	<i>Abbreviation for</i> length(which(x))
----	--

Description

lw takes a logical vector, x, and returns length(which(x)).

lw takes a logical vector, x, and returns length(which(x)).

Usage

```
lw(x)
```

```
lw(x)
```

Arguments

x A logical

Value

The length of the TRUE values in x

The length of the TRUE values in x

See Also

[length, which](#)

[length, which](#)

Other tcpl abbreviations: [is.odd\(\)](#), [lu\(\)](#), [sink.reset\(\)](#)

Other tcpl abbreviations: [is.odd\(\)](#), [lu\(\)](#), [sink.reset\(\)](#)

mc1

Perform level 1 multiple-concentration processing

Description

mc1 loads level 0 data from the tcpl database for the given id and performs level 1 multiple-concentration processing. The processed data is then loaded into the mc1 table and all subsequent data is deleted with [tcplCascade](#). See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the [tcplRun](#) wrapper function.

Usage

```
mc1(ac, wr = FALSE)
```

Arguments

ac Integer of length 1, assay component id (acid) for processing.

wr Logical, whether the processed data should be written to the tcpl database

Details

Level 1 processing includes defining the concentration and replicate index, cndx and repi, respectively.

Value

A boolean of length 1, indicating the success of the processing, or when 'wr' is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a data.table containing the processed data

See Also

Other multiple-concentration: [mc2\(\)](#), [mc3\(\)](#), [mc4\(\)](#), [mc5\(\)](#), [mc6\(\)](#)

mc2

Perform level 2 multiple-concentration processing

Description

mc2 loads level 1 data from the tcpl database for the given id and performs level 2 multiple-concentration processing. The processed data is then loaded into the mc2 table and all subsequent data is deleted with [tcplCascade](#). See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the [tcplRun](#) wrapper function.

Usage

```
mc2(ac, wr = FALSE)
```

Arguments

ac	Integer of length 1, assay component id (acid) for processing.
wr	Logical, whether the processed data should be written to the tcpl database

Details

Level 2 multiple-concentration processing includes defining the corrected value, cval, based on the correction methods listed in the mc2_acid and mc2_methods tables.

Value

A boolean of length 1, indicating the success of the processing, or when 'wr' is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a data.table containing the processed data

See Also

[Method functions](#), [MC2_Methods](#)

Other multiple-concentration: [mc1\(\)](#), [mc3\(\)](#), [mc4\(\)](#), [mc5\(\)](#), [mc6\(\)](#)

Description

mc2_mthds returns a list of correction/transformation functions to be used during level 2 multiple-concentration processing.

Usage

```
mc2_mthds()
```

Details

The functions contained in the list returned by mc2_mthds return a list of expressions to be executed in the mc2 (not exported) function environment. The functions are described here for reference purposes, The mc2_mthds function is not exported, nor is it intended for use.

All available methods are described in the Available Methods section, listed by the function/method name.

Value

A list functions

Available Methods

More information about the level 2 multiple-concentration processing is available in the package vignette, "Pipeline_Overview."

log2 Take the logarithm of cval with the base 2.

log10 Take the logarithm of cval with the base 10.

rmneg Remove entries where cval is less than 0.

rmzero Remove entries where cval is 0.

mult25 Multiply cval by 25.

mult100 Multiply cval by 100.

negshift Shift cval by subtracting out the minimum of cval and adding 1, such that the new minimum of cval is 1.

mult25 Multiply cval by 2.5.

mult3 Multiply cval by 3.

mult6 Multiply cval by 6.

Note

This function is not exported and is not intended to be used by the user.

See Also

[mc2](#), [Method functions](#) to query what methods get applied to each acid

mc3

Perform level 3 multiple-concentration processing

Description

mc3 loads level 2 data from the tcpl database for the given id and performs level 3 multiple-concentration processing. The processed data is then loaded into the mc3 table and all subsequent data is deleted with [tcplCascade](#). See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the [tcplRun](#) wrapper function.

Usage

```
mc3(ac, wr = FALSE)
```

Arguments

ac	Integer of length 1, assay component id (acid) for processing.
wr	Logical, whether the processed data should be written to the tcpl database

Details

Level 3 multiple-concentration processing includes mapping assay component to assay endpoint, duplicating the data when the assay component has multiple assay endpoints, and any normalization of the data. Data normalization based on methods listed in mc3_aeid and mc3_methods tables.

Value

A boolean of length 1, indicating the success of the processing, or when 'wr' is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a data.table containing the processed data

See Also

[Method functions](#), [MC3_Methods](#)

Other multiple-concentration: [mc1\(\)](#), [mc2\(\)](#), [mc4\(\)](#), [mc5\(\)](#), [mc6\(\)](#)

Description

mc3_mthds returns a list of normalization methods to be used during level 3 multiple-concentration processing.

Usage

```
mc3_mthds()
```

Details

The functions contained in the list returned by mc3_mthds take 'aeids' (a numeric vector of aeid values) and returns a list of expressions to be executed in the mc3 (not exported) function environment. The functions are described here for reference purposes, The mc3_mthds function is not exported, nor is it intended for use.

All available methods are described in the Available Methods section, listed by the type of function and the function/method name.

Value

A list of functions

Available Methods

The methods are broken into three types, based on what fields they define. Different methods are used to define "bval" (the baseline value), "pval" (the positive control value), and "resp" (the final response value).

Although it does not say so specifically in each description, all methods are applied by aeid.

More information about the level 3 multiple-concentration processing is available in the package vignette, "Pipeline_Overview."

bval Methods:

bval.apid.nwlls.med Calculate bval as the median of cval for wells with wllt equal to "n," by apid.

bval.apid.lowconc.med Calculate bval as the median of cval for wells with wllt equal to "t" and cndx equal to 1 or 2, by apid.

bval.apid.twlls.med Calculate bval as the median of cval for wells with wllt equal to "t," by apid.

bval.apid.tn.med Calculate bval as the median of cval for wells with wllt equal to "t" or "n," by apid.

bval.apid.nwllslowconc.med Calculate bval as the median of cval for wells with wllt equal to "n" or wells with wllt equal to "t" and cndx equal to 1 or 2, by apid.

bval.spid.lowconc.med Calculate bval as the median of cval for wells with wllt equal to "t" and cndx equal to 1, 2, or 3, by spid.

bval.apid.nwllstcwlslowconc.med Calculate bval as the median of cval for wells with wllt equal to "n" or cndx equal to 1 or 2 and wllt equal to "t" or "c" by apid.

pval Methods:

pval.apid.pwlls.med Calculate pval as the median of cval for wells with wllt equal to "p," by apid.

pval.apid.mwlls.med Calculate pval as the median of cval for wells with wllt equal to "m," by apid.

pval.apid.medpcbyconc.max First calculate the median of cval for wells with wllt equal to "p" or "c," by wllt, conc, and apid. Then calculate pval as the maximum of the calculated medians, by apid.

pval.apid.medpcbyconc.min First calculate the median of cval for wells with wllt equal to "p" or "c," by wllt, conc, and apid. Then calculate pval as the minimum of the calculated medians, by apid.

pval.apid.medncbyconc.min First calculate the median of cval for wells with wllt equal to "m" or "o," by wllt, conc, and apid. Then calculate pval as the minimum of the calculated medians, by apid.

pval.apid.pmv.min First calculate the median of cval for wells with wllt equal to "p," "m," or "v," by wllt, conc, and apid. Then calculate pval as the minimum of the calculated medians, by apid.

pval.apid.pmv.max First calculate the median of cval for wells with wllt equal to "p," "m," or "v," by wllt, conc, and apid. Then calculate pval as the maximum of the calculated medians, by apid.

pval.apid.f.max First calculate the median of cval for wells with wllt equal to "f," by wllt, conc, and apid. Then calculate pval as the maximum of the calculated medians, by apid.

pval.apid.f.min First calculate the median of cval for wells with wllt equal to "f," by wllt, conc, and apid. Then calculate pval as the minimum of the calculated medians, by apid.

pval.apid.p.max First calculate the median of cval for wells with wllt equal to "p," by wllt, conc, and apid. Then calculate pval as the maximum of the calculated medians, by apid.

pval.apid.p.min First calculate the median of cval for wells with wllt equal to "p," by wllt, conc, and apid. Then calculate pval as the minimum of the calculated medians, by apid.

pval.apid.v.min First calculate the median of cval for wells with wllt equal to "v," by wllt, conc, and apid. Then calculate pval as the minimum of the calculated medians, by apid.

pval.zero Define pval as 0.

resp Methods:

resp.pc Calculate resp as $\frac{cval-bval}{pval-bval}100$.

resp.pc.pval.cor Calculate resp as $\frac{cval-bval}{pval}100$.

resp.fc Calculate resp as $cval/bval$.

resp.logfc Calculate resp as $cval - bval$.

resp.log2 Take the logarithm of resp with base 2.

resp.mult25 Multiply resp by 25.

resp.scale.mad.log2fc Multiply resp by the scale factor $\frac{\log_2(1.2)}{3bmad}$.

resp.scale.quant.log2fc Determine the maximum response md where $md = \text{abs}(1\text{st centile} - 50\text{th centile})$ or $\text{abs}(99\text{th centile} - 50\text{th centile})$, whichever is greater. Scale the response such that 20 percent of md equals $\log_2(1.2)$.

- resp.multneg1** Multiply resp by -1.
- resp.shiftneg.3bmad** Shift all resp values less than $-3*bmad$ to 0.
- resp.shiftneg.6bmad** Shift all resp values less than $-6*bmad$ to 0.
- resp.shiftneg.10bmad** Shift all resp values less than $-10*bmad$ to 0.
- resp.blineshift.3bmad.repi** Shift resp values with the `blineshift` function by `repi`, where the window (`wndw`) is $3*bmad$.
- resp.blineshift.50.repi** Shift resp values with the `blineshift` function by `repi`, where the window (`wndw`) is 50.
- resp.blineshift.3bmad.spid** Shift resp values with the `blineshift` function by `spid`, where the window (`wndw`) is $3*bmad$.
- resp.blineshift.50.spid** Shift resp values with the `blineshift` function by `spid`, where the window (`wndw`) is 50.
- none** Do no normalization; make resp equal to `cval`.

Note

This function is not exported and is not intended to be used by the user.

See Also

[mc3](#), [Method functions](#) to query what methods get applied to each `aeid`

mc4

*Perform level 4 multiple-concentration processing***Description**

`mc4` loads level 3 data from the `tcpl` database for the given `id` and performs level 4 multiple-concentration processing. The processed data is then loaded into the `mc4` table and all subsequent data is deleted with `tcplCascade`. See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the `tcplRun` wrapper function.

Usage

```
mc4(ae, wr = FALSE)
```

Arguments

<code>ae</code>	Integer of length 1, assay endpoint id (<code>aeid</code>) for processing.
<code>wr</code>	Logical, whether the processed data should be written to the <code>tcpl</code> database

Details

Level 4 multiple-concentration modeling takes the dose-response data for chemical-assay pairs, and fits three models to the data: constant, hill, and gain-loss. For more information about the models see [Models](#). When a chemical has more than one sample, the function fits each sample separately.

Value

A boolean of length 1, indicating the success of the processing, or when 'wr' is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a data.table containing the processed data

See Also

[tcp1Fit, Models](#)

Other multiple-concentration: [mc1\(\)](#), [mc2\(\)](#), [mc3\(\)](#), [mc5\(\)](#), [mc6\(\)](#)

MC4_Methods

List of level 4 multiple-concentration methods for calculating bmad

Description

mc4_mthds returns a list of methods to be used during level 4 multiple-concentration processing for calculating bmad

Usage

```
mc4_mthds()
```

Details

The functions contained in the list returned by mc4_mthds take 'aeids' (a numeric vector of aeid values) and returns a list of expressions to be executed in the mc4 (not exported) function environment. The functions are described here for reference purposes, The mc4_mthds function is not exported, nor is it intended for use.

All available methods are described in the Available Methods section, listed by the type of function and the function/method name.

Value

A list of functions

Available Methods

Although it does not say so specifically in each description, all methods are applied by aeid.

More information about the level 4 multiple-concentration processing is available in the package vignette, "Pipeline_Overview."

bmad.aeid.lowconc.twells bmad based on two lowest concentration of treatment wells

bmad.aeid.lowconc.nwells bmad based on two lowest concentration of nwells

Note

This function is not exported and is not intended to be used by the user.

See Also

[mc4](#), [Method functions](#) to query what methods get applied to each aeid

mc5

Perform level 5 multiple-concentration processing

Description

mc5 loads level 4 data from the tcpl database for the given id and performs level 5 multiple-concentration processing. The processed data is then loaded into the mc5 table and all subsequent data is deleted with [tcplCascade](#). See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the [tcplRun](#) wrapper function.

Arguments

ae	Integer of length 1, assay endpoint id (aeid) for processing.
wr	Logical, whether the processed data should be written to the tcpl database

Details

Level 5 multiple-concentration hit-calling uses the fit parameters and the activity cutoff methods from mc5_aeid and mc5_methods to make an activity call and identify the winning model for each fit.

Value

A boolean of length 1, indicating the success of the processing, or when 'wr' is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a data.table containing the processed data

See Also

[Method functions](#), [MC5_Methods](#)

Other multiple-concentration: [mc1\(\)](#), [mc2\(\)](#), [mc3\(\)](#), [mc4\(\)](#), [mc6\(\)](#)

MC5_Methods

Load list of level 5 multiple-concentration cutoff methods

Description

mc5_mthds returns a list of additional activity cutoff methods to be used during level 5 multiple-concentration processing.

Usage

```
mc5_mthds(ae)
```

Arguments

ae Integer of length 1, the assay endpoint id

Value

A list of functions

Available Methods

More information about the level 5 multiple-concentration processing is available in the package vignette, "Pipeline_Overview."

bmad3 Add a cutoff value of 3*bmad.

pc20 Add a cutoff value of 20.

log2_1.2 Add a cutoff value of log2(1.2).

log10_1.2 Add a cutoff value of log10(1.2).

bmad5 Add a cutoff value of 5*bmad.

bmad6 Add a cutoff value of 6*bmad.

bmad10 Add a cutoff value of 10*bmad.

log2_2 Add a cutoff value of log2(2).

log10_2 Add a cutoff value of log10(2).

neglog2_0.88 Add a cutoff value of -1*log2(0.88).

coff_2.32 Add a cutoff value of 2.32.

See Also

[mc5](#), [Method functions](#) to query what methods get applied to each acid

mc6	<i>Perform level 6 multiple-concentration processing</i>
-----	--

Description

mc6 loads level 5 data from the tcpl database for the given id and performs level 6 multiple-concentration processing. The processed data is then loaded into the mc6 table and all subsequent data is deleted with [tcplCascade](#). See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the [tcplRun](#) wrapper function.

Usage

```
mc6(ae, wr = FALSE)
```

Arguments

ae	Integer of length 1, assay endpoint id (aeid) for processing.
wr	Logical, whether the processed data should be written to the tcpl database

Details

Level 6 multiple-concentration flagging uses both the plate level concentration-response data and the modeled parameters to flag potential false positives and false negative results.

Value

A boolean of length 1, indicating the success of the processing, or when 'wr' is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a data.table containing the processed data

See Also

[Method functions](#), [MC6_Methods](#)

Other multiple-concentration: [mc1\(\)](#), [mc2\(\)](#), [mc3\(\)](#), [mc4\(\)](#), [mc5\(\)](#)

MC6_Methods

*Load list of level 6 multiple-concentration flag methods***Description**

mc6_mthds returns a list of flag methods to be used during level 6 multiple-concentration processing.

Usage

```
mc6_mthds()
```

Value

A list functions

Available Methods

More information about the level 6 multiple-concentration processing is available in the package vignette, "Pipeline_Overview."

singlept.hit.high The singlept.hit.high flag identifies concentration series where the median response was greater than 3*bmad only at the highest tested concentration and the series had an active hit-call.

singlept.hit.mid The singlept.hit.mid flag identifies concentration series where the median response was greater than 3*bmad at only one concentration (not the highest tested concentration) and the series had an active hit-call.

multipoint.neg The multipoint.neg flag identifies concentration series with response medians greater than 3*bmad at multiple concentrations and an inactive hit-call.

gnls.lowconc The gnls.lowconc flag identifies concentration series where the gain-loss model won, the gain AC50 is less than the minimum tested concentration, and the loss AC50 is less than the mean tested concentration.

noise The noise flag attempts to identify noisy concentration series by flagging series where the root mean square error for the series is greater than the cutoff for the assay endpoint.

border.hit The border.hit flag identifies active concentration series where the top parameter of the winning model was less than or equal to 1.2*cut-off or the the activity probability was less than 0.9.

border.miss The border.miss flag identifies inactive concentration series where either the Hill or gain-loss top parameter was greater than or equal to 0.8*cut-off and the activity probability was greater than 0.5.

overfit.hit The overfit.hit flag recalculates the model winner after applying a small sample correction factor to the AIC values. If the hit-call would be changed after applying the small sample correction factor the series is flagged. Series with less than 5 concentrations where the hill model won and series with less than 7 concentrations where the gain-loss model won are automatically flagged.

efficacy.50 The efficacy.50 flag identifies concentration series with efficacy values (either the modeled top parameter for the winning model or the maximum median response) are less than 50 for percent activity data or $\log_2(1.5)$ for fold induction data

modlga.lowconc The modlga.lowconc flag identifies concentration series with modl_ga (AC50) values less than the minimum tested concentration.

See Also

[mc6](#), [Method functions](#) to query what methods get applied to each acid

mcdat	<i>A subset of ToxCast data showing changes in the activity of the intracellular estrogen receptor.</i>
-------	---

Description

The example dataset is used to illustrate how the user can pipeline multiple-concentration data from chemical screening using tcplite.

Usage

```
mcdat
```

Format

A data frame with 14183 rows and 10 variables:

spid sample ID
apid assay plate ID
rowi well-plate row number
coli well-plate column number
wllt well type
wllq well quality
conc concentration in micromolar
rval raw assay component readout value
srfc source file containing the data
acsn assay component source name

Source

ToxCast database

`mc_vignette`*List with multi-concentration data for the vignette*

Description

This dataset is a list with 6 data.tables (mc0,mc1,mc2,mc3,mc4,mc5).

Usage`mc_vignette`**Format**

1. **mc0** A data frame with 78 rows and 18 columns containing level 0 formatted raw data.
 - spid** Sample ID
 - chid** Unique chemical ID number for tcpl
 - casn** Chemical Abstract Service(CAS) number
 - chnm** Chemical name
 - dsstox_substance_id** Chemical-specific DTXSID
 - code** CAS number compressed into numeric string
 - acid** Assay Component ID
 - acnm** Assay Component Name
 - m0id** Level 0 (mc0) ID
 - apid** Assay plate ID
 - rowi** Row Index
 - coli** Column Index
 - wllt** Well Type
 - wllq** Well Quality (0 or 1)
 - conc** Concentration in micromolar
 - rval** Raw assay component readout value
 - srcf** Source file containing the raw data
 - conc_unit** Concentration Units
2. **mc1** A data frame with 78 rows and 21 columns containing level 1 replicate and concentration level indicated data.
 - spid** Sample ID
 - chid** Unique chemical ID number for tcpl
 - casn** Chemical Abstract Service(CAS) number
 - chnm** Chemical name
 - dsstox_substance_id** Chemical-specific DTXSID
 - code** CAS number compressed into numeric string
 - acid** Assay Component ID
 - acnm** Assay Component Name

- m0id** Level 0 (mc0) ID
 - m1id** Level 1 (mc1) ID
 - apid** Assay plate ID
 - rowi** Row Index
 - coli** Column Index
 - wllt** Well Type
 - wllq** Well Quality (0 or 1)
 - conc** Concentration in micromolar
 - rval** Raw assay component readout value
 - ndcx** Concentration index defined by ranking the unique concentrations, with the lowest concentration starting at 1.
 - repi** Temporary replicate ID is defined, the data are scanned from top to bottom and increment the replicate index every time a replicate ID is duplicated
 - srcf** Source file containing the raw data
 - conc_unit** Concentration Units
3. **mc2** A data frame with 78 rows and 20 columns containing level 2 assay component-specific corrections.
- spid** Sample ID
 - chid** Unique chemical ID number for tcpl
 - casn** Chemical Abstract Service(CAS) number
 - chnm** Chemical name
 - dsstox_substance_id** Chemical-specific DTXSID
 - code** CAS number compressed into numeric string
 - acid** Assay Component ID
 - acnm** Assay Component Name
 - m0id** Level 0 (mc0) ID
 - m1id** Level 1 (mc1) ID
 - m2id** Level 2 (mc2) ID
 - apid** Assay plate ID
 - rowi** Row Index
 - coli** Column Index
 - wllt** Well Type
 - conc** Concentration in micromolar
 - cval** Corrected Value
 - ndcx** Concentration index defined by ranking the unique concentrations, with the lowest concentration starting at 1.
 - repi** Temporary replicate ID is defined, the data are scanned from top to bottom and increment the replicate index every time a replicate ID is duplicated
 - conc_unit** Concentration Units
4. **mc3** A data frame with 78 rows and 22 columns containing level 3 assay endpoint normalized data.
- spid** Sample ID

- chid** Unique chemical ID number for tcpl
 - casn** Chemical Abstract Service(CAS) number
 - chnm** Chemical name
 - dsstox_substance_id** Chemical-specific DTXSID
 - code** CAS number compressed into numeric string
 - aeid** Assay Component Endpoint ID
 - aenm** Assay endpoint name (i.e., assay_component_endpoint_name)
 - m0id** Level 0 (mc0) ID
 - m1id** Level 1 (mc1) ID
 - m2id** Level 2 (mc2) ID
 - m3id** Level 3 (mc3) ID
 - logc** Log base 10 concentration
 - resp** Normalized response value
 - cndx** Concentration index defined by ranking the unique concentrations, with the lowest concentration starting at 1.
 - wllt** Well Type
 - apid** Assay plate ID
 - rowi** Row Index
 - coli** Column Index
 - repi** Temporary replicate ID is defined, the data are scanned from top to bottom and increment the replicate index every time a replicate ID is duplicated
 - resp_unit** Response Units
 - conc_unit** Concentration Units
5. **mc4** A data frame with 5 rows and 149 columns containing level 4 concentration-response fitting data (all fits).
- spid** Sample ID
 - chid** Unique chemical ID number for tcpl
 - casn** Chemical Abstract Service(CAS) number
 - chnm** Chemical name
 - dsstox_substance_id** Chemical-specific DTXSID
 - code** CAS number compressed into numeric string
 - aeid** Assay Component Endpoint ID
 - aenm** Assay endpoint name (i.e., assay_component_endpoint_name)
 - m4id** Level 4 (mc4) ID
 - bmad** The median absolute deviation of all treatment wells (default option) or blank wells
 - resp_max** Maximum observed response
 - resp_min** Minimum observed response
 - max_mean** Maximum mean response
 - max_mean_conc** Concentration of the maximum mean response
 - max_med** Maximum median response
 - max_med_conc** Concentration of the maximum median response
 - logc_max** Maximum concentration on the log scale

logc_min Minimum concentration on the log scale
nconc The total number of concentration groups
npts Total number of observed responses (i.e. data points in the concentration series)
nrep Number of replicates in concentration groups
nmed_gtbl The number of median responses greater than 3BMAD
cnst_success Success indicator for the Constant model; 1 if the optimization was successful, otherwise 0
cnst_aic Akaike Information Criteria (AIC) for the Constant model
cnst_rme Root mean square error for the Constant model
cnst_er Error term for the Constant model
hill_success Success indicator for the Hill model; 1 if the optimization was successful, otherwise 0
hill_aic Akaike Information Criteria (AIC) for the Hill model
hill_cov Success indicator for the Hill model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0
hill_rme Root mean square error for the Hill model
hill_tp The top parameter indicating the maximal estimated response
hill_ga The gain parameter for the Hill model, gain AC50
hill_p The power parameter for the Hill model
hill_er Error term for the Hill model
hill_tp_sd Standard deviation of the Hill model top parameter
hill_ga_sd Standard deviation of the Hill model gain parameter
hill_p_sd Standard deviation of the Hill model power parameter
hill_er_sd Standard deviation of the Hill model error term
hill_top The maximal response on the resulting Hill model fit
hill_ac50 Concentration at 50% of the maximal response on the Hill model fit
gnls_success Success indicator for the Gain-loss model; 1 if the optimization was successful, otherwise 0
gnls_aic Akaike Information Criteria (AIC) for the Gain-loss model
gnls_cov Success indicator for the Gain-loss model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0
gnls_rme Root mean square error for the Gain-loss model
gnls_tp The top parameter indicating the maximal estimated response
gnls_ga The gain parameter for the Gain-loss model, gain AC50
gnls_p The gain power parameter for the Gain-loss model
gnls_la The loss parameter for the Gain-loss model, loss AC50
gnls_q The loss power parameter for the Gain-loss model
gnls_er Error term for the Gain-loss model
gnls_tp_sd Standard deviation of the Gain-loss model top parameter
gnls_ga_sd Standard deviation of the Gain-loss model gain parameter
gnls_p_sd Standard deviation of the Gain-loss model gain power parameter
gnls_la_sd Standard deviation of the Gain-loss model loss parameter
gnls_q_sd Standard deviation of the Gain-loss model loss power parameter

gnls_er_sd Standard deviation of the Gain-loss model error term

gnls_top The maximal response on the resulting Gain-loss model fit

gnls_ac50 Concentration at 50% of the maximal response on the Gain-loss model fit, gain AC50

gnls_ac50_loss Concentration at 50% of the maximal response on the Gain-loss model fit, loss AC50

poly1_success Success indicator for the Polynomial 1 model; 1 if the optimization was successful, otherwise 0

poly1_aic Akaike Information Criteria (AIC) for the Polynomial 1 model

poly1_cov Success indicator for the Polynomial 1 model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0

poly1_rme Root mean square error for the Polynomial 1 model

poly1_a The y-scale parameter for the Polynomial 1 model

poly1_er Error term for the Polynomial 1 model

poly1_a_sd Standard deviation of the Polynomial 1 model y-scale parameter

poly1_er_sd Standard deviation of the Polynomial 1 model error term

poly1_top The maximal response on the resulting Polynomial 1 model fit

poly1_ac50 Concentration at 50% of the maximal response on the Polynomial 1 model fit

poly2_success Success indicator for the Polynomial 2 model; 1 if the optimization was successful, otherwise 0

poly2_aic Akaike Information Criteria (AIC) for the Polynomial 2 model

poly2_cov Success indicator for the Polynomial 2 model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0

poly2_rme Root mean square error for the Polynomial 2 model

poly2_a The y-scale parameter for the Polynomial 2 model

poly2_b The x-scale parameter for the Polynomial 2 model

poly2_er Error term for the Polynomial 2 model

poly2_a_sd Standard deviation of the Polynomial 2 model y-scale parameter

poly2_b_sd Standard deviation of the Polynomial 2 model x-scale parameter

poly2_er_sd Standard deviation of the Polynomial 2 model error term

poly2_top The maximal response on the resulting Polynomial 2 model fit

poly2_ac50 Concentration at 50% of the maximal response on the Polynomial 2 model fit

pow_success Success indicator for the Power model; 1 if the optimization was successful, otherwise 0

pow_aic Akaike Information Criteria (AIC) for the Power model

pow_cov Success indicator for the Power model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0

pow_rme Root mean square error for the Power model

pow_a The y-scale parameter for the Power model

pow_p The power parameter for the Power model

pow_er Error term for the Power model

pow_a_sd Standard deviation of the Power model y-scale parameter

pow_p_sd Standard deviation of the Power model power parameter

pow_er_sd Standard deviation of the Power model error term

pow_top The maximal response on the resulting Power model fit

pow_ac50 Concentration at 50% of the maximal response on the Power model fit

exp2_success Success indicator for the Exponential 2 model; 1 if the optimization was successful, otherwise 0

exp2_aic Akaike Information Criteria (AIC) for the Exponential 2 model

exp2_cov Success indicator for the Exponential 2 model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0

exp2_rme Root mean square error for the Exponential 2 model

exp2_a The y-scale parameter for the Exponential 2 model

exp2_b The x-scale parameter for the Exponential 2 model

exp2_er Error term for the Exponential 2 model

exp2_a_sd Standard deviation of the Exponential 2 model y-scale parameter

exp2_b_sd Standard deviation of the Exponential 2 model x-scale parameter

exp2_er_sd Standard deviation of the Exponential 2 model error term

exp2_top The maximal response on the resulting Exponential 2 model fit

exp2_ac50 Concentration at 50% of the maximal response on the Exponential 2 model fit

exp3_success Success indicator for the Exponential 3 model; 1 if the optimization was successful, otherwise 0

exp3_aic Akaike Information Criteria (AIC) for the Exponential 3 model

exp3_cov Success indicator for the Exponential 3 model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0

exp3_rme Root mean square error for the Exponential 3 model

exp3_a The y-scale parameter for the Exponential 3 model

exp3_b The x-scale parameter for the Exponential 3 model

exp3_p The power parameter for the Exponential 3 model

exp3_er Error term for the Exponential 3 model

exp3_a_sd Standard deviation of the Exponential 3 model y-scale parameter

exp3_b_sd Standard deviation of the Exponential 3 model x-scale parameter

exp3_p_sd Standard deviation of the Exponential 3 model power parameter

exp3_er_sd Standard deviation of the Exponential 3 model error term

exp3_top The maximal response on the resulting Exponential 3 model fit

exp3_ac50 Concentration at 50% of the maximal response on the Exponential 3 model fit

exp4_success Success indicator for the Exponential 4 model; 1 if the optimization was successful, otherwise 0

exp4_aic Akaike Information Criteria (AIC) for the Exponential 4 model

exp4_cov Success indicator for the Exponential 4 model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0

exp4_rme Root mean square error for the Exponential 4 model

exp4_tp The top parameter indicating the maximal estimated response

exp4_ga The gain parameter for the Exponential 4 model, gain AC50

exp4_er Error term for the Exponential 4 model

exp4_tp_sd Standard deviation of the Exponential 4 model top parameter

- exp4_ga_sd** Standard deviation of the Exponential 4 model gain parameter
 - exp4_er_sd** Standard deviation of the Exponential 4 model error term
 - exp4_top** The maximal response on the resulting Exponential 4 model fit
 - exp4_ac50** Concentration at 50% of the maximal response on the Exponential 4 model fit
 - exp5_success** Success indicator for the Exponential 5 model; 1 if the optimization was successful, otherwise 0
 - exp5_aic** Akaike Information Criteria (AIC) for the Exponential 5 model
 - exp5_cov** Success indicator for the Exponential 5 model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0
 - exp5_rme** Root mean square error for the Exponential 5 model
 - exp5_tp** The top parameter indicating the maximal estimated response
 - exp5_ga** The gain parameter for the Exponential 5 model, gain AC50
 - exp5_p** The power parameter for the Exponential 5 model
 - exp5_er** Error term for the Exponential 5 model
 - exp5_tp_sd** Standard deviation of the Exponential 5 model top parameter
 - exp5_ga_sd** Standard deviation of the Exponential 5 model gain parameter
 - exp5_p_sd** Standard deviation of the Exponential 5 model power parameter
 - exp5_er_sd** Standard deviation of the Exponential 5 model error term
 - exp5_top** The maximal response on the resulting Exponential 5 model fit
 - exp5_ac50** Concentration at 50% of the maximal response on the Exponential 5 model fit
 - all_onesd** Standard deviation of the baseline response for all models
 - all_bmed** Median noise estimation of the baseline response for all models
 - resp_unit** Response Units
 - conc_unit** Concentration Units
6. **mc5** A data frame with 5 rows and 54 columns containing level 5 best curve-fit and hitcall data.
- spid** Sample ID
 - chid** Unique chemical ID number for tcpl
 - casn** Chemical Abstract Service(CAS) number
 - chnm** Chemical name
 - dsstox_substance_id** Chemical-specific DTXSID
 - code** CAS number compressed into numeric string
 - aecid** Assay Component Endpoint ID
 - aenm** Assay endpoint name (i.e., assay_component_endpoint_name)
 - m5id** Level 5 (mc5) ID
 - m4id** Level 4 (mc4) ID
 - bmad** The median absolute deviation of all treatment wells (default option) or blank wells
 - resp_max** Maximum observed response
 - resp_min** Minimum observed response
 - max_mean** Maximum mean response
 - max_mean_conc** Concentration of the maximum mean response
 - max_med** Maximum median response

max_med_conc Concentration of the maximum median response
logc_max Maximum concentration on the log scale
logc_min Minimum concentration on the log scale
nconc The total number of concentration groups
npts Total number of observed responses (i.e. data points in the concentration series)
nrep Number of replicates in concentration groups
nmed_gtbl The number of median responses greater than 3BMAD
hitc Hitcall
modl Best model fit from tcplFit2 curve-fitting
fitc Fit category
coff Cutoff
top_over_cutoff Ratio of the top of the best model fit curve and the cutoff
rmse Root mean squared error
a The y-scale parameter for poly1, poly2, pow, exp2, or exp3 model
er Error term
bmr Benchmark response
bmdl Lower 95% confidence bound on the benchmark dose/concentration estimate
caikwt Akaike Information Criteria weight of constant model relative to the best model fit
mll Maximum log-likelihood of the best model fit
hitcall Continuous hitcall
ac50 Concentration where 50% of the maximal response occurs - if 'modl' is the Hill or Gain-loss model this is for the "gain" side of the response
top The maximal response on the best model curve fit - i.e. top of the curve fit
ac5 Concentration where 5% of the maximal response occurs
ac10 Concentration where 10% of the maximal response occurs
ac20 Concentration where 20% of the maximal response occurs
acc Concentration where the efficacy cutoff response occurs
ac1sd Concentration where one standard deviation of the background response occurs
bmd Benchmark response/concentration estimate - concentration where the benchmark response occurs
bmdu Upper 95% confidence bound on the benchmark dose/concentration estimate
tp The top curve parameter for the exp4, exp5, hill, or gnls model
ga The gain parameter for the hill or gnls model - gain AC50
p The power parameter for the pow, exp3, exp5, gnls, or hill model - for gnls this is the gain power parameter
q The loss power parameter for the gnls model
la The loss parameter for the gnls model, loss AC50
ac50_loss Concentration where 50% of the maximal response occurs - if 'modl' is the Hill or Gain-loss model this is for the "loss" side of the response
b The x-scale parameter for poly2, exp2, or exp3 model
resp_unit Response Units
conc_unit Concentration Units

Method functions *Functions for managing processing methods*

Description

These functions are used to manage which methods are used to process data. They include methods for assigning, clearing, and loading the assigned methods. Also, `tcplMthdList` lists the available methods.

Usage

```
tcplMthdAssign(lvl, id, mthd_id, ordr = NULL, type)
tcplMthdClear(lvl, id, mthd_id = NULL, type)
tcplMthdList(lvl, type = "mc")
tcplMthdLoad(lvl, id = NULL, type = "mc")
```

Arguments

<code>lvl</code>	Integer of length 1, the method level
<code>id</code>	Integer, the assay component or assay endpoint id(s)
<code>mthd_id</code>	Integer, the method id(s)
<code>ordr</code>	Integer, the order in which to execute the analysis methods, must be the same length as <code>mthd_id</code> , does not apply to levels 5 or 6
<code>type</code>	Character of length 1, the data type, "sc" or "mc"

Details

`tcplMthdLoad` loads the assigned methods for the given level and ID(s). Similarly, `tcplMthdList` displays the available methods for the given level. These two functions do not make any changes to the database.

Unlike the `-Load` and `-List` functions, the `-Assign` and `-Clear` functions alter the database and trigger a delete cascade. `tcplMthdAssign` assigns methods to the given ID(s), and `tcplMthdClear` removes methods. In addition to the method ID (`'mthd_id'`), assigning methods at some levels require an order (`'ordr'`). The `'ordr'` parameter is necessary to allow progression of methods at level one for single-concentration processing, and levels two and three for multiple-concentration processing. More information about method assignments and the delete cascade are available in the package vignette.

Examples

```
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcplConfList()
```

```

tcplConfDefault()

## tcplListMthd allows the user to display the available methods for
## a given level and data type
head(tcplMthdList(lvl = 2, type = "mc"))

## tcplLoadMthd shows which methods are assigned for the given ID, level,
## and data type. Here we will show how to register, load, and clear methods
## using an acid not in the example database. Note: There is no check for
## whether an ID exists before assigning/clearing methods.
tcplMthdLoad(lvl = 2, id = 55, type = "mc")

## Not run:
## ACID 55 does not have any methods. Assign methods from the list above.
tcplMthdAssign(lvl = 2,
               id = 55,
               mthd_id = c(3, 4, 2),
               ordr = 1:3,
               type = "mc")
## Method assignment can be done for multiple assays, too.
tcplMthdAssign(lvl = 2,
               id = 53:54,
               mthd_id = c(3, 4, 2),
               ordr = 1:3,
               type = "mc")

## Cleanup example method assignments
tcplMthdClear(lvl = 2, id = 53:55, type = "mc")

## End(Not run)
## Reset configuration
options(conf_store)

```

Models

Model objective functions

Description

These functions take in the dose-response data and the model parameters, and return a likelihood value. They are intended to be optimized using `constrOptim` in the `tcplFit` function.

Usage

```

tcplObjCnst(p, resp)

tcplObjGnls(p, lconc, resp)

tcplObjHill(p, lconc, resp)

tcplObjCnst(p, resp)

```

```
tcplObjGnls(p, lconc, resp)
```

```
tcplObjHill(p, lconc, resp)
```

Arguments

p	Numeric, the parameter values. See details for more information.
resp	Numeric, the response values
lconc	Numeric, the log10 concentration values

Details

These functions produce an estimated value based on the model and given parameters for each observation. Those estimated values are then used with the observed values and a scale term to calculate the log-likelihood.

Let $t(z, \nu)$ be the Student's t-distribution with ν degrees of freedom, y_i be the observed response at the i^{th} observation, and μ_i be the estimated response at the i^{th} observation. We calculate z_i as:

$$z_i = \frac{y_i - \mu_i}{e^\sigma}$$

where σ is the scale term. Then the log-likelihood is:

$$\sum_{i=1}^n [\ln(t(z_i, 4)) - \sigma]$$

Where n is the number of observations.

Value

The log-likelihood.

Constant Model (cnst)

tcplObjCnst calculates the likelihood for a constant model at 0. The only parameter passed to tcplObjCnst by p is the scale term σ . The constant model value μ_i for the i^{th} observation is given by:

$$\mu_i = 0$$

tcplObjCnst calculates the likelihood for a constant model at 0. The only parameter passed to tcplObjCnst by p is the scale term σ . The constant model value μ_i for the i^{th} observation is given by:

$$\mu_i = 0$$

Gain-Loss Model (gnls)

tcp1objGnls calculates the likelihood for a 5 parameter model as the product of two Hill models with the same top and both bottoms equal to 0. The parameters passed to tcp1objGnls by p are (in order) top (tp), gain log AC50 (ga), gain hill coefficient (gw), loss log AC50 la , loss hill coefficient lw , and the scale term (σ). The gain-loss model value μ_i for the i^{th} observation is given by:

$$g_i = \frac{1}{1 + 10^{(ga-x_i)gw}}$$

$$l_i = \frac{1}{1 + 10^{(x_i-la)lw}}$$

$$\mu_i = tp(g_i)(l_i)$$

where x_i is the log concentration for the i^{th} observation.

tcp1objGnls calculates the likelihood for a 5 parameter model as the product of two Hill models with the same top and both bottoms equal to 0. The parameters passed to tcp1objGnls by p are (in order) top (tp), gain log AC50 (ga), gain hill coefficient (gw), loss log AC50 la , loss hill coefficient lw , and the scale term (σ). The gain-loss model value μ_i for the i^{th} observation is given by:

$$g_i = \frac{1}{1 + 10^{(ga-x_i)gw}}$$

$$l_i = \frac{1}{1 + 10^{(x_i-la)lw}}$$

$$\mu_i = tp(g_i)(l_i)$$

where x_i is the log concentration for the i^{th} observation.

Hill Model (hill)

tcp1objHill calculates the likelihood for a 3 parameter Hill model with the bottom equal to 0. The parameters passed to tcp1objHill by p are (in order) top (tp), log AC50 (ga), hill coefficient (gw), and the scale term (σ). The hill model value μ_i for the i^{th} observation is given by:

$$\mu_i = \frac{tp}{1 + 10^{(ga-x_i)gw}}$$

where x_i is the log concentration for the i^{th} observation.

tcp1objHill calculates the likelihood for a 3 parameter Hill model with the bottom equal to 0. The parameters passed to tcp1objHill by p are (in order) top (tp), log AC50 (ga), hill coefficient (gw), and the scale term (σ). The hill model value μ_i for the i^{th} observation is given by:

$$\mu_i = \frac{tp}{1 + 10^{(ga-x_i)gw}}$$

where x_i is the log concentration for the i^{th} observation.

Description

These functions send a query to the given database, and are the access point for all tcpl functions that query or update the tcpl database.

Usage

```
tcplQuery(
  query,
  db = getOption("TCPL_DB"),
  drvr = getOption("TCPL_DRVR"),
  tbl = NULL
)

tcplSendQuery(
  query,
  db = getOption("TCPL_DB"),
  drvr = getOption("TCPL_DRVR"),
  tbl = NULL,
  delete = F
)
```

Arguments

query	Character of length 1, the query string
db	Character of length 1, the name of the tcpl database
drvr	Character of length 1, which database driver to use
tbl	Tables to be read queried
delete	Logical of length 1, execute delete on queried table

Details

Currently, the tcpl package only supports the "MySQL" and "tcplLite" database drivers.

tcplQuery returns a data.table object with the query results. tcplSendQuery sends a query, but does not fetch any results, and returns 'TRUE' or the error message given by the database.

Examples

```
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcplConfList()
TCPLlite <- file.path(system.file(package = "tcpl"), "example")
```



```
tcplConf(db = TCPLlite, user = NA, host = NA, drvr = "tcplLite")

tcplQuery("SELECT 'Hello World';")

## When using tcplLite, name of table must be passed into tcplQuery
if (conf_store$TCPL_DRVR == 'MySQL') {
  tcplQuery("SELECT * FROM assay;")
} else {
  tcplQuery("SELECT * FROM assay;", tbl='assay')
}

## Reset configuration
options(conf_store)
```

Register/update annotation

Functions for registering & updating annotation information

Description

These functions are used to register and update the chemical and assay annotation information.

Usage

```
tcplRegister(what, flds)

tcplUpdate(what, id, flds)
```

Arguments

what	Character of length 1, the name of the ID to register or update
flds	Named list, the other fields and their values
id	Integer, the ID value(s) to update

Details

These functions are used to populate the tcpl database with the necessary annotation information to complete the processing. As shown in the package vignette, the package requires some information about the samples and assays before data can be loaded into the tcpl database.

Depending on what is being registered, different information is required. The following table lists the fields that can be registered/updated by these functions, and the minimal fields required for registering a new ID. (The database table affected is in parentheses.)

- asid (assay_source): assay_source_name
- aid (assay): asid, assay_name, assay_footprint
- acid (assay_component): aid, assay_component_name

- `acid` (`assay_component_endpoint`): `acid`, `assay_component_endpoint_name`, `normalized_data_type`
- `acsn` (`assay_component_map`): `acid`, `acsn`
- `spid` (`sample`): `spid`, `chid`
- `chid` (`chemical`): `chid`, `casn`
- `clib` (`chemical_library`): `chid`, `clib`

Note: The functions accept the abbreviated forms of the names, ie. "aenm" rather than the full "assay_component_endpoint_name." More information about the registration process and all of the fields is available in the vignette.

Examples

```
## Not run:
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcplConfList()
tcplConfDefault()

## Load current ASID information
tcplLoadAsid()

## Register a new assay source
tcplRegister(what = "asid", flds = list(asnm = "example_asid"))

## Show the newly registered ASID
tcplLoadAsid(add.fld = "assay_source_desc")

## Notice that the newly created ASID does not have an assay_source_desc.
## The field could have been defined during the registration process, but
## can also be updated using tcplUpdate
i1 <- tcplLoadAsid()[asnm == "example_asid", asid]
tcplUpdate(what = "asid",
           id = i1,
           flds = list(assay_source_desc = "example asid description"))
tcplLoadAsid(add.fld = "assay_source_desc")

## Remove the created ASID. Note: Manually deleting primary keys can cause
## serious database problems and should not generally be done.

## If using the tcplLite DRVR, must specify table name
if (conf_store$TCPL_DRVR == 'MySQL') {
  tcplSendQuery(paste0("DELETE FROM assay_source WHERE asid = ", i1, ";"))
} else {
  qy <- paste0("SELECT * FROM assay_source WHERE NOT asid = ", i1, ";")
  tcplSendQuery(qy, tbl='assay_source', delete=TRUE)
}

## Reset configuration
options(conf_store)
```

```
## End(Not run)
```

registerMthd	<i>Add a new analysis method</i>
--------------	----------------------------------

Description

registerMthd registers a new analysis method to the tcpl databases.

Usage

```
registerMthd(lvl, mthd, desc, nldr = 0L, type)
```

Arguments

lvl	Integer of length 1, the level for the analysis method
mthd	Character, the name of the method
desc	Character, same length as mthd, the method description
nldr	Integer, 0 or 1, 1 if the method requires loading the dose- response data
type	Character of length 1, the data type, "sc" or "mc"

Details

'mthd' must match a corresponding function name in the functions that load the methods, ie. mc2_mthds. 'nldr' only applies to level 6 methods.

sc1	<i>Perform level 1 single-concentration processing</i>
-----	--

Description

sc1 loads level 0 data from the tcpl database for the given id and performs level 1 single-concentration processing. The processed data is then loaded into the sc1 table and all subsequent data is deleted with [tcplCascade](#). See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the [tcplRun](#) wrapper function.

Usage

```
sc1(ac, wr = FALSE)
```

Arguments

ac	Integer of length 1, assay component id (acid) for processing.
wr	Logical, whether the processed data should be written to the tcpl database

Details

Level 1 single-concentration processing includes mapping assay component to assay endpoint, duplicating the data when the assay component has multiple assay endpoints, and any normalization of the data. Data normalization based on methods listed in `sc1_aeid` and `sc1_methods` tables.

Value

A boolean of length 1, indicating the success of the processing, or when 'wr' is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a `data.table` containing the processed data

See Also

[Method functions, SC1_Methods](#)

Other single-concentration: [sc2\(\)](#)

SC1_Methods

List of level 1 single-concentration normalization functions

Description

`sc1_mthds` returns a list of functions to be used during level 1 single-concentration processing.

Usage

```
sc1_mthds()
```

Details

The functions contained in the list returned by `sc1_mthds` return a list of expressions to be executed in the `sc2` (not exported) function environment. The functions are described here for reference purposes, The `sc1_mthds` function is not exported, nor is it intended for use.

All available methods are described in the Available Methods section, listed by the function/method name.

Value

A list functions

Available Methods

The methods are broken into three types, based on what fields they define. Different methods are used to define "bval" (the baseline value), "pval" (the positive control value), and "resp" (the final response value).

Although it does not say so specifically in each description, all methods are applied by acid.

More information about the level 3 single-concentration processing is available in the package vignette, "Pipeline_Overview."

bval Methods:

bval.apid.nwlls.med Calculate bval as the median of rval for wells with wllt equal to "n," by apid.

bval.apid.twlls.med Calculate bval as the median of rval for wells with wllt equal to "t," by apid.

bval.apid.tn.med Calculate bval as the median of rval for wells with wllt equal to "t" or "n," by apid.

pval Methods:

pval.apid.pwlls.med Calculate pval as the median of rval for wells with wllt equal to "p," by apid.

pval.apid.mwlls.med Calculate pval as the median of rval for wells with wllt equal to "m," by apid.

pval.apid.medpcbyconc.max First calculate the median of rval for wells with wllt equal to "p" or "c," by wllt, conc, and apid. Then calculate pval as the maximum of the calculated medians, by apid.

pval.apid.medpcbyconc.min First calculate the median of rval for wells with wllt equal to "p" or "c," by wllt, conc, and apid. Then calculate pval as the minimum of the calculated medians, by apid.

pval.apid.medncbyconc.min First calculate the median of rval for wells with wllt equal to "m" or "o," by wllt, conc, and apid. Then calculate pval as the minimum of the calculated medians, by apid.

pval.zero Define pval as 0.

resp Methods:

resp.pc Calculate resp as $\frac{rval - bval}{pval - bval} 100$.

resp.fc Calculate resp as $rval / bval$.

resp.logfc Calculate resp as $rval - bval$.

resp.log2 Take the logarithm of resp with base 2.

resp.multneg1 Multiply resp by -1.

none Do no normalization; make resp equal to rval.

Note

This function is not exported and is not intended to be used by the user.

See Also

[sc1, Method functions](#) to query what methods get applied to each acid

sc2	<i>Perform level 2 single-concentration processing</i>
-----	--

Description

sc2 loads level 1 data from the tcpl database for the given id and performs level 2 single-concentration processing. The processed data is then loaded into the sc2 table and all subsequent data is deleted with [tcplCascade](#). See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the [tcplRun](#) wrapper function.

Usage

```
sc2(ae, wr = FALSE)
```

Arguments

ae	Integer of length 1, assay endpoint id (aeid) for processing.
wr	Logical, whether the processed data should be written to the tcpl database

Details

Level 2 single-concentration processing defines the bmad value, and uses the activity cutoff methods from `sc2_aeid` and `sc2_methods` to make an activity call.

Value

A boolean of length 1, indicating the success of the processing, or when 'wr' is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a `data.table` containing the processed data

See Also

[Method functions](#), [SC2_Methods](#)

Other single-concentration: [sc1\(\)](#)

Description

sc2_mthds returns a list of functions to be used during level 2 single-concentration processing.

Usage

```
sc2_mthds()
```

Details

The functions contained in the list returned by sc2_mthds return a list of expressions to be executed in the sc2 (not exported) function environment. The functions are described here for reference purposes, The sc2_mthds function is not exported, nor is it intended for use.

All available methods are described in the Available Methods section, listed by the function/method name.

Value

A list functions

Available Methods

More information about the level 2 single-concentration processing is available in the package vignette, "Pipeline_Overview."

bmad3 Add a cutoff value of 3*bmad.

pc20 Add a cutoff value of 20.

log2_1.2 Add a cutoff value of log2(1.2).

log10_1.2 Add a cutoff value of log10(1.2).

bmad5 Add a cutoff value of 5*bmad.

bmad6 Add a cutoff value of 6*bmad.

bmad10 Add a cutoff value of 10*bmad.

pc30orbmad3 Add a cutoff value of either 30 or 3*bmad, whichever is less.

ow_bmad_nwells Overwrite method to calculate bmad based on nwells for acid.

bmad2 Add a cutoff value of 2*bmad.

bmad1 Add a cutoff value of 1*bmad.

Note

This function is not exported and is not intended to be used by the user.

See Also

[sc2, Method functions](#) to query what methods get applied to each acid

sccdat	<i>A subset of ToxCast data showing changes in transcription factor activity for multiple targets.</i>
--------	--

Description

The example dataset is used to illustrate how the user can pipeline single-concentration data from chemical screening using tcplite.

Usage

sccdat

Format

A data frame with 320 rows and 10 variables:

spid sample ID
apid assay plate ID
rowi well-plate row number (N/A)
coli well-plate column number (N/A)
wllt well type (N/A)
wllq well quality (N/A)
conc concentration in micromolar
rval raw assay component readout value
srcf source file containing the data
acsn assay component source name

Source

ToxCast database

sc_vignette	<i>List with single-concentration data for the vignette</i>
-------------	---

Description

This dataset is a list with 3 data.tables (sc0,sc1,sc2).

Usage

sc_vignette

Format

1. **sc0** A data frame with 10 rows and 18 columns containing level 0 formatted raw data.

spid Sample ID
chid Unique chemical ID number for tcpl
casn Chemical Abstract Service(CAS) number
chnm Chemical name
dsstox_substance_id Chemical-specific DTXSID
code CAS number compressed into numeric string
acid Assay Component ID
acnm Assay Component Name
s0id Level 0 (sc0) ID
apid Assay plate ID
rowi Row Index
coli Column Index
wllt Well Type
wllq Well Quality (0 or 1)
conc Concentration in micromolar
rval Raw assay component readout value
srcf Source file containing the raw data
conc_unit Concentration Units

2. **sc1** A data frame with 10 rows and 20 columns containing level 1 normalized data.

spid Sample ID
chid Unique chemical ID number for tcpl
casn Chemical Abstract Service(CAS) number
chnm Chemical name
dsstox_substance_id Chemical-specific DTXSID
code CAS number compressed into numeric string
acid Assay Component Endpoint ID
aenm Assay endpoint name (i.e., assay_component_endpoint_name)
acid Assay Component ID
acnm Assay Component Name
s0id Level 0 (sc0) ID
s1id Level 1 (sc1) ID
apid Assay plate ID
rowi Row Index
coli Column Index
wllt Well Type
logc Log base 10 concentration
resp Normalized response value
resp_unit Response Units
conc_unit Concentration Units

3. **sc2** A data frame with 10 rows and 15 columns containing level 2 efficacy/hit designation data.

spid Sample ID
chid Unique chemical ID number for tcpl
casn Chemical Abstract Service(CAS) number
chnm Chemical name
dsstox_substance_id Chemical-specific DTXSID
code CAS number compressed into numeric string
aeid Assay Component Endpoint ID
aenm Assay endpoint name (i.e., assay_component_endpoint_name)
s2id Level 2 (sc2) ID
bmad The median absolute deviation of all treatment wells (default option) or blank wells
max_med Maximum median response
hitc Hitcall
coff Cutoff
resp_unit Response Units
conc_unit Concentration Units

sink.reset

Reset all sinks

Description

sink.reset resets all sinks and returns all output to the console.

Usage

```
sink.reset()
```

Details

sink.reset identifies all sinks with sink.number then returns all output and messages back to the console.

See Also

[sink](#), [sink.number](#)

Other tcpl abbreviations: [is.odd\(\)](#), [lu\(\)](#), [lw\(\)](#)

tcplAddModel	<i>Draw a tcpl Model onto an existing plot</i>
--------------	--

Description

tcplAddModel draws a a line for one of the tcpl Models (see [Models](#) for more information) onto an existing plot.

Usage

```
tcplAddModel(pars, modl = NULL, adj = NULL, ...)
```

Arguments

pars	List of parameters from level 4 or 5 output
modl	Character of length 1, the model to plot: 'cnst,' 'hill,' or 'gnls'
adj	Numeric of length 1, an adjustment factor, see details for more information
...	Additional arguments passed to curve

Details

tcplAddModel draws the model line assuming the x-axis represents log base 10 concentration.

If modl is NULL, the function checks pars\$modl and will return an error if pars\$modl is also NULL.

adj is intended to scale the models, so that models with different response units can be visualized on a single plot. The recommended value for adl is $1/(3 \cdot \text{bmad})$ for level 4 data and $1/\text{coff}$ for level 5 data. If adj is NULL the function will check pars\$adj and set adj to 1 if pars\$adj is also NULL.

See Also

[Models](#), [tcplPlotFits](#)

Examples

```
## Create some dummy data to plot
logc <- 1:10
r1 <- sapply(logc, tcplHillVal, ga = 5, tp = 50, gw = 0.5)
r2 <- log2(sapply(logc, tcplHillVal, ga = 4, tp = 30, gw = 0.5))
p1 <- tcplFit(logc = logc, resp = r1, bmad = 10)
p2 <- tcplFit(logc = logc, resp = r2, bmad = log2(1.5))

## In the dummy data above, the two plots are on very different scales
plot(r1 ~ logc, pch = 16, ylab = "raw response")
tcplAddModel(pars = p1, modl = "hill")
points(r2 ~ logc)
tcplAddModel(pars = p2, modl = "hill", lty = "dashed")
```

```
## To visualize the two curves on the same plot for comparison, we can
## scale the values to the bmad, such that a scaled response of 1 will equal
## the bmad for each curve.
plot(r1/10 ~ logc, pch = 16, ylab = "scaled response")
tcplAddModel(pars = p1, modl = "hill", adj = 1/10)
points(r2/log2(5) ~ logc)
tcplAddModel(pars = p2, modl = "hill", adj = 1/log2(5), lty = "dashed")
```

tcplAICProb

Calculate the AIC probabilities

Description

tcplAICProb Calculates the probability that the model best represents the data based on the AIC value for each model.

Usage

```
tcplAICProb(...)
```

Arguments

... Numeric vectors of AIC values

Details

The function takes vectors of AIC values. Each vector represents the model AIC values for multiple observation sets. Each vector must contain the same number and order of observation sets. The calculation assumes every possible model is accounted for, and the results should be interpreted accordingly.

Value

A vector of probability values for each model given, as a list.

See Also

[tcplFit](#), [AIC](#) for more information about AIC values.

Examples

```
## Returns the probability for each model, given models with AIC values
## ranging from 80 to 100
tcplAICProb(80, 85, 90, 95, 100)

## Also works for vectors
m1 <- c(95, 195, 300) ## model 1 for three different observations
```

```
m2 <- c(100, 200, 295) ## model 2 for three different observations
tcplAICProb(m1, m2)
```

tcplAppend	<i>Append rows to a table</i>
------------	-------------------------------

Description

tcplAppend takes a data.table (dat) and appends the data.table into a database table.

Usage

```
tcplAppend(dat, tbl, db, lvl = NULL)
```

Arguments

dat	data.table, the data to append to a table
tbl	Character of length 1, the table to append to
db	Character of length 1, the database containing tbl
lvl	Usually Integer to indicate what level to auto-increment

Note

This function is not exported and not intended to be used by the user.

tcplCascade	<i>Do a cascading delete on tcpl screening data</i>
-------------	---

Description

tcplCascade deletes the data for the given id(s) starting at the processing level given. The delete will cascade through all subsequent tables.

Usage

```
tcplCascade(lvl, type, id)
```

Arguments

lvl	Integer of length 1, the first level to delete from
type	Character of length 1, the data type, "sc" or "mc"
id	Integer, the id(s) to delete. See details for more information.

Details

The data type can be either 'mc' for multiple concentration data, or 'sc' for single concentration data. Multiple concentration data will be loaded into the level tables, whereas the single concentration will be loaded into the single tables.

If lvl is less than 3, id is interpreted as acid(s) and if lvl is greater than or equal to 3, id is interpreted as aeid(s).

Note

This function is not exported and not intended to be used by the user.

tcp1Code2CASN	<i>Convert chemical code to CAS Registry Number</i>
---------------	---

Description

tcp1Code2CASN takes a code and converts it CAS Registry Number.

Usage

```
tcp1Code2CASN(code)
```

Arguments

code	Character of length 1, a chemical code
------	--

Details

The function checks for the validity of the CAS Registry Number. Also, the ToxCast data includes chemicals for which there is no CASRN. The convention for these chemicals is to give them a CASRN as NOCAS_chid; the code for these compounds is CNOCASchid. The function handles the NOCAS compounds as they are stored in the database, as shown in the example below.

Value

A CAS Registry Number.

Examples

```
tcp1Code2CASN("C80057")
tcp1Code2CASN("C09812420") ## Invalid CASRN will give a warning
tcp1Code2CASN("CNOCAS0015") ## The underscore is reinserted for NOCAS codes
```

tcplCytoPt	<i>Calculate the cytotoxicity point based on the "burst" endpoints</i>
------------	--

Description

tcplCytoPt calculates the cytotoxicity point and average cytotoxicity distribution based on the activity in the "burst" assay endpoints.

Usage

```
tcplCytoPt(
  chid = NULL,
  aeid = NULL,
  flag = TRUE,
  min.test = TRUE,
  default.pt = 3
)
```

Arguments

chid	Integer, chemical ID values to subset on
aeid	Integer, assay endpoint ID values to override the "burst assay" definitions
flag	Integer, mc6_mthd_id values to be passed to tcplSubsetChid
min.test	Integer or Boolean, the number of tested assay endpoints required for a chemical to be used in calculating the "global MAD."
default.pt	Numeric of length 1, the default cytotoxicity point value

Details

tcplCytoPt provides estimates for chemical-specific cytotoxicity distributions (more information available in the vignette.) Before calculating the cytotoxicity distributions, the level 5 data is subsetted by the [tcplSubsetChid](#) function.

The 'chid' parameter specifies a subset of chemicals to use in the calculations, given by chemical ID (chid). The 'aeid' parameter specifies which assays to use in calculating the cytotoxicity point and distribution. By default tcplCytoPt will use all available chemicals and the assay endpoints defined by the 'burst_assay' field in the "assay_component_endpoint" table. The examples show how to identify the "burst" endpoints.

tcplCytoPt returns the cytotoxicity point (the AC50 values of the active "burst" endpoints), the corresponding MAD, and the global MAD (median of the calculated MAD values). Not every chemical must be tested in every "burst" endpoint. The 'min.test' parameter allows the user to specify a minimum number of tested assay endpoints as a requirement for MAD values to be included in the global MAD calculation. For example, suppose the user supplies 10 "burst" assays. The user can choose to require a chemical to be tested in at least 5 of those assays for its MAD value to be included in the global MAD calculation. Having chemicals with many less "burst" endpoints tested may inflate or deflate the global MAD calculation. By default (values of TRUE or NULL),

tcplCytoPt requires a chemical to be tested in at least 80% of the given "burst" assays. The user can also provide 'min.test' values of FALSE (indicating to include all MAD values), or a number (indicating a specific number of endpoints).

Chemicals without at least 2 active "burst" assays do not have a MAD value, and the cytotoxicity point is defined by the 'default.pt' parameter. The default value for 'default.pt' is 3.

The resulting data.table has the following fields:

1. "chid" – The chemical ID.
2. "code" – The chemical code.
3. "chnm" – The chemical name.
4. "casn" – The chemical CASRN.
5. "med" – The median of the "burst" endpoint log(AC50) ("modl_ga" in the level 5 output) values.
6. "mad" – The MAD of the "burst" endpoint log(AC50) values.
7. "ntst" – The number of "burst" endpoints tested.
8. "nhit" – The number of active "burst" endpoints.
9. "use_global_mad" – TRUE/FALSE, whether the mad value was used in the global MAD calculation.
10. "global_mad" – The median of the "mad" values where "use_global_mad" is TRUE.
11. "cyto_pt" – The cytotoxicity point, or the value in "med" when "nhit" is at least 2.
12. "cyto_pt_um" – $10^{\text{cyto_pt}}$
13. "lower_bnd_um" – $10^{\text{cyto_pt} - 3\text{global_mad}}$

Value

A data.table with the cytotoxicity distribution for each chemical. The definition of the field names are listed under "details."

Examples

```
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcplConfList()
tcplConfDefault()

## Can only calculate the cytotox burst if using the MySQL database and
## TCPL_DRVR == 'MySQL'

if (getOption("TCPL_DRVR") == "MySQL") {

  ## Load the "burst" endpoints -- none are defined in the example dataset
  tcplLoadAeid(fld = "burst_assay", val = 1)

  ## Calculate the cytotoxicity distributions using both example endpoints
  tcplCytoPt(aeid = 1:2)
```



```

## The above example does not calculate a global MAD, because no chemical
## hit both endpoints. (This makes sense, because both endpoints are
## derived from one component, where one endpoint is activity in the
## up direction, and the other is activity in the down direction.)
## Note, the cyto_pt is also 3 for all chemicals, because the function
## requires at least two endpoints to calculate a cytotoxicity point. If
## the user wishes to use one assay, this function is not necessary.

## Changing 'default.pt' will change cyto_pt in the resulting data.table
tcplCytoPt(aeid = 1:2, default.pt = 6)
}

## Reset configuration
options(conf_store)

```

tcpldbStats

Get summary statistics for the database

Description

tcpldbStats takes a string(*type*) and an optional parameter(*val*) to return the summary statistics on the entire tcplLite database. When *type* = "all" the *val* is ignored. the function returns the number of distinct *spid* and *aeids* in the database at each level. When *type* = "aeid", the *val* parameter has to be a valid *aeid* in the database. The function returns a table consisting of the number of distinct *spids* at each level of processing for the *aeid* given in '*val*'. When *type* = "spid", the *val* parameter has to be a valid *spid* in the database. The function returns a table consisting of the number of distinct *aeids* at each level of processing for the given *spid* in '*val*'.

Usage

```
tcpldbStats(type = "all", val = NULL)
```

Arguments

<i>type</i>	String either "all", "aeid" or "spid"
<i>val</i>	integer if <i>type</i> = "aeid" , string if <i>type</i> = "spid"

tcplDelete

Delete rows from tcpl databases

Description

tcplDelete deletes rows from the given table and database.

Usage

```
tcplDelete(tbl, fld, val, db)
```

Arguments

tbl	Character, length 1, the table to delete from
fld	Character, the field(s) to query on
val	List, vectors of values for each field to query on. Must be in the same order as 'fld'.
db	Character, the database containing the table

Note

This function is not exported and not intended to be used by the user.

See Also

[tcplSendQuery](#)

tcplFit

Fit the data with the constant, hill, and gain-loss models

Description

tcplFit fits the constant, hill, and gain-loss models to the given data and returns some summary statistics and the fit parameters in a list.

Usage

```
tcplFit(  
  logc,  
  resp,  
  bmad,  
  force.fit = FALSE,  
  bidirectional = FALSE,  
  verbose = FALSE,  
  ...  
)
```

Arguments

logc	Numeric, log concentration values
resp	Numeric, normalized response values
bmad	Numeric, the baseline median absolute deviation for the entire assay
force.fit	Logical, TRUE indicates to attempt fitting every concentration series

bidirectional Boolean If TRUE, bidirectional negative data before fitting (default=FALSE)
 The original version of the code required the data to start at small values and rise, so that negative curves had to be bidirectionalped outside the function, and TOP was always positive. Setting bidirectional to TRUE allows both rising and falling curves

verbose Boolean If TRUE print warning messages

... Any other data to be included in list output.

Details

when at least one median value is greater than 3*bmad.

Value

List of summary values and fit parameters for the given data.

See Also

[tcplObjCnst](#), [tcplObjHill](#), [tcplObjGnls](#), [constrOptim](#)

Examples

```

logc <- 1:10
resp <- sapply(1:10, tcplHillVal, ga = 5, tp = 50, gw = 0.5)
params <- tcplFit(logc = logc, resp = resp, bmad = 10)
plot(resp ~ logc)
tcplAddModel(pars = params, modl = "hill")

```

tcplFit2	<i>tcpl Wrapper for tcplfit2_core including additional calculations to fit into new schema</i>
----------	--

Description

tcpl Wrapper for tcplfit2_core including additional calculations to fit into new schema

Usage

```

tcplFit2(
  dat,
  fitmodels = c("cnst", "hill", "gnls", "poly1", "poly2", "pow", "exp2", "exp3", "exp4",
    "exp5"),
  bmed = NULL,
  bidirectional = TRUE
)

```

Arguments

dat	output from level 3 processing
fitmodels	list of the models that should be fit with the data
bmed	baseline value, typically should be 0
bidirectional	boolean, default is TRUE (bidirectional fitting)

Value

Data.table with an additional column fitparams that includes all of the fitting parameters

tcplFit2_nest	<i>Nest dataframe into a list that is readable by tcplfit2</i>
---------------	--

Description

Nest dataframe into a list that is readable by tcplfit2

Usage

```
tcplFit2_nest(dat)
```

Arguments

dat	a dataframe that has all of the fitting parameters in the style of tcplloaddata
-----	---

Value

a list of fitting parameters that can be consumed by tcplfit2

tcplFit2_unnest	<i>Unnest tcplfit2 parameters into a dataframe</i>
-----------------	--

Description

Unnest tcplfit2 parameters into a dataframe

Usage

```
tcplFit2_unnest(output)
```

Arguments

output	list of output from tcplfit2
--------	------------------------------

Value

list of parameters unnested and compiled into a dataframe

tcplGetAeid	<i>get Aeid for endpoint name</i>
-------------	-----------------------------------

Description

tcplGetAeid takes a string(name) and finds the assay component endpoint names that match the string and the aeid associated with those names. The function performs a regular expression like matching for strings in the assay component endpoint name column in the assay component endpoint table.

Usage

```
tcplGetAeid(name)
```

Arguments

name A string that will be matched to the assay component endpoint name

Examples

```
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcplConfList()
tcplConfExample()

## Search for aenm (assay name) case insensitive
tcplGetAeid("TOX21")
tcplGetAeid("tox21")

## Reset configuration
options(conf_store)
```

tcplggplot	<i>tcplggplot</i>
------------	-------------------

Description

tcplggplot

Usage

```
tcplggplot(dat, lvl = 5, verbose = FALSE)
```

Arguments

dat	data table with all required conc/resp data
lvl	integer level of data that should be plotted level 4 - all fit models level 5 - all fit models and winning model with hitcall level 6 - include all flags
verbose	boolean should plotting include table of values next to the plot

Value

A ggplot object or grob with accompanied table depending on verbose option

tcp1Hit2	<i>Hitcalling with tcp1fit2</i>
----------	---------------------------------

Description

Hitcalling with tcp1fit2

Usage

```
tcp1Hit2(mc4, coff)
```

Arguments

mc4	data.table with level 4 data
coff	cutoff value for hitcalling

Value

Data.table with key value pairs of hitcalling parameters

tcpListFlds	<i>Load the field names for a table</i>
-------------	---

Description

tcpListFlds loads the column names for the given table and database.

Usage

```
tcpListFlds(tbl, db = getOption("TCPL_DB"))
```

Arguments

tbl	Character of length 1, the tcp database table
db	Character of length 1, the tcp database

Details

This function can be particularly useful in defining the 'fld' param in the tcplLoad- functions.

Value

A string of field names for the given table.

Examples

```
## Gives the fields in the mc1 table
tcplListFlds("mc1")
```

tcplLoadChem	<i>Load sample/chemical information</i>
--------------	---

Description

tcplLoadChem queries the tcpl database and returns the chemical information for the given field and values.

Usage

```
tcplLoadChem(field = NULL, val = NULL, exact = TRUE, include.spid = TRUE)
```

Arguments

field	Character of length 1, the field to query on
val	Vector of values to subset on
exact	Logical, should chemical names be considered exact?
include.spid	Logical, should spid be included?

Details

The 'field' parameter is named differently from the 'fld' parameter seen in other functions because it only takes one input.

In the MySQL environment the user should be able to give partial chemical name strings, to find chemicals with similar names. For example, setting 'val' to "phenol" when 'field' is "chnm" and 'exact' is FALSE might pull up the chemicals "Bisphenol A" and "4-Butylphenol". More technically, setting 'exact' to FALSE passes the string in 'val' to an RLIKE statement within the MySQL query.

Value

A data.table with the chemical information for the given parameters

Examples

```

## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcplConfList()
tcplConfExample()

## Passing no parameters gives all of the registered chemicals with their
## sample IDs
tcplLoadChem()

## Or the user can exclude spid and get a unique list of chemicals
tcplLoadChem(include.spid = FALSE)

## In addition, the user can retrieve only the registered chemicals from the chemical table
tcplLoadChem(field = 'chem.only')

## Other examples:
tcplLoadChem(field = "chnm", val = "Bisphenol A")
tcplLoadChem(field = "chid", val = 20182)

## Reset configuration
options(conf_store)

```

tcplLoadClib

Load chemical library information

Description

tcplLoadClib queries the tcpl databases and returns information about the chemical library.

Usage

```
tcplLoadClib(field = NULL, val = NULL)
```

Arguments

field	Character of length 1, 'chid' or 'clib', whether to search by chemical id (chid), or chemical library (clib)
val	The values to query on

Details

Chemicals are stored in different libraries by chemical ID. Therefore, it is not possible to delineate samples with the same chemical ID into two distinct chemical libraries. However, it is possible for a chemical ID to belong to more than one (or no) chemical libraries.

When chemicals belong to more than one library, the chemical is listed multiple times (one for each distinct library).

Value

A data.table with the chemical library information for the given parameters.

Examples

```
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcplConfList()
tcplConfExample()

## Passing no parameters gives all of the chemical ISs that have a chemical
## library registered
clib <- tcplLoadClib()

## Notice there are more rows in tcplLoadClib than in tcplLoadChem,
## indicating some chemicals must belong to more than library.
chem <- tcplLoadChem(include.spid = FALSE)
nrow(chem)
nrow(clib)

## It is possible that some chemicals do not have a chemical library
## registered, although this is not the case in the example data.
all(chem$chid %in% clib$chid)

## Show the unique chemical libraries
clib[, unique(clib)]

## Specifying a chemical library will not show what other libraries a
## chemical might belong to.
tcplLoadClib(field = "clib", val = "TOXCAST")
tcplLoadClib(field = "chid", val = 20182)

## Reset configuration
options(conf_store)
```

tcplLoadConcUnit *Load concentration units for assay endpoints*

Description

tcplLoadUnit queries the tcpl databases and returns a data.table with the concentration units for the given assay endpoint ids (spid).

Usage

```
tcplLoadConcUnit(spid)
```

Arguments

spid Integer, assay endpoint ids

Value

A data.table containing level 3 correction methods for the given spids.

See Also

[tcpQuery](#), [data.table](#)

tcpLoadData	<i>Load tcp data</i>
-------------	----------------------

Description

tcpLoadData queries the tcp databases and returns a data.table with data for the given level and data type.

Usage

```
tcpLoadData(lvl, fld = NULL, val = NULL, type = "mc", add.fld = NULL)
```

Arguments

lvl Integer of length 1, the level of data to load

fld Character, the field(s) to query on

val List, vectors of values for each field to query on. Must be in the same order as 'fld'.

type Character of length 1, the data type, "sc" or "mc"

add.fld Boolean if true we want to return the additional parameters fit with tcpfit2

Details

The data type can be either 'mc' for multiple concentration data, or 'sc' for single concentration data. Multiple concentration data will be loaded into the 'mc' tables, whereas the single concentration will be loaded into the 'sc' tables.

Setting 'lvl' to "agg" will return an aggregate table containing the m4id with the concentration-response data and m3id to map back to well-level information.

Leaving fld NULL will return all data.

Valid fld inputs are based on the data level and type:

type	lvl	Queried tables
sc	0	sc0
sc	1	sc0, sc1

sc	agg	sc1, sc2_agg
sc	2	sc2
mc	0	mc0
mc	1	mc0, mc1
mc	2	mc0, mc1, mc2
mc	3	mc0, mc1, mc3
mc	agg	mc3, mc4_agg
mc	4	mc4
mc	5	mc4, mc5
mc	6	mc4, mc6
mc	7	mc4, mc7

Value

A data.table containing data for the given fields.

See Also

[tcplQuery](#), [data.table](#)

Examples

```
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcplConfList()
tcplConfExample()

## Load all of level 0 for multiple-concentration data, note 'mc' is the
## default value for type
tcplLoadData(lvl = 0)

## Load all of level 1 for single-concentration
tcplLoadData(lvl = 1, type = "sc")

## List the fields available for level 1, coming from tables mc0 and mc1
tcplListFlds(tbl = "mc0")
tcplListFlds(tbl = "mc1")

## Load level 0 data where the well type is "t" and the concentration
## index is 3 or 4
tcplLoadData(lvl = 1, fld = c("wllt", "cndx"), val = list("t", c(3:4)))

## Reset configuration
options(conf_store)
```

Description

tcpLoadUnit queries the tcp databases and returns a data.table with the response units for the given assay endpoint ids (aeid).

Usage

```
tcpLoadUnit(aeid)
```

Arguments

aeid Integer, assay endpoint ids

Value

A data.table containing level 3 correction methods for the given aeids.

See Also

[tcpQuery](#), [data.table](#)

tcpLvlCount

Load tcp level counts

Description

tcpLvlCount queries the tcp databases and returns a data frame with count totals for the given levels and data type.

Usage

```
tcpLvlCount(lvls = NULL, type = "mc")
```

Arguments

lvls Integer or list of Integers, The levels of data to load

type Character of length 1, the data type, "sc" or "mc"

Details

The data type can be either 'mc' for mutiple concentration data, or 'sc' for single concentration data.

Leaving lvls NULL will return all data.

Value

A data.table containing data for the given fields.

See Also

[tcplQuery](#), [data.table](#)

Examples

```
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcplConfList()
TCPLlite <- file.path(system.file(package = "tcpl"), "example")
tcplConf(db = TCPLlite, user = NA, host = NA, drvr = "tcplLite")

## Get all counts for level 1 for multiple-concentration
tcplLvlCount(lvls = 1)

## Not run:
## Get all counts for levels 4 through 7 for multiple-concentration
tcplLvlCount(lvls = 4:7)

## Get all counts for multiple-concentration data, note 'mc' is the
## default value for type
tcplLvlCount()

## End(Not run)

## Reset configuration
options(conf_store)
```

tcplMakeAeidMultiPlts *Create a .pdf with all dose-response plots for a given aeid, 6 per page*

Description

tcplMakeAeidMultiPlts Create a .pdf with all dose-response plots for a given aeid

Usage

```
tcplMakeAeidMultiPlts(
  aeid,
  lvl = 4L,
  fname = NULL,
  odir = getwd(),
  clib = NULL,
  hitc.all = TRUE
)
```

Arguments

aeid	Integer of length 1, the assay endpoint id
lvl	Integer of length 1, the data level to use (4-7)
fname	Character, the filename
odir	The directory to save the .pdf file in
clib	Character, the chemical library to subset on, see tcplLoadClib for more information.
hitc.all	If FALSE, only plots with hitc==1 will be displayed

Details

tcplMakeAeidMultiPlts provides a wrapper for [tcplMultiplot](#), allowing the user to produce PDFs with the curve plots without having to separately load all of the data and establish the PDF device.

If 'fname' is NULL, a default name is given by concatenating together assay information.

tcplMakeAeidPlts	<i>Create a .pdf with dose-response plots</i>
------------------	---

Description

tcplMakeAeidPlts creates a .pdf file with the dose-response plots for the given aeid.

Usage

```
tcplMakeAeidPlts(
  aeid,
  compare = F,
  lvl = 4L,
  fname = NULL,
  odir = getwd(),
  ordr.fitc = TRUE,
  clib = NULL,
  cnst = NULL
)
```

Arguments

aeid	Integer of length 1 or 2, the assay endpoint id
compare	Boolean to for comparison of aeids if length(aeid)>1
lvl	Integer of length 1, the data level to use (4-7). Only level 5-6 valid for compare aeids.
fname	Character, the filename
odir	The directory to save the .pdf file in

ordr.fitc	Logical, should the fits be ordered by fit category?
clib	Character, the chemical library to subset on, see
cnst	Constant hline to draw on plot tcplLoadClib for more information.

Details

tcplMakeAeidPlts provides a wrapper for [tcplPlotFits](#), allowing the user to produce PDFs with the curve plots without having to separately load all of the data and establish the PDF device.

If 'fname' is NULL, a default name is given by concatenating together assay information.

Note, the default value for ordr.fitc is TRUE in tcplMakeAeidPlts, but FALSE in tcplPlotFits

Note, only level 5 or level 6 is valid for comparing 2 aeids.

Examples

```
## Not run:
## Will produce the same result as the example for tcplPlotFits
tcplMakeAeidPlts(aeid = 1, lvl = 6, ordr.fitc = FALSE)

## End(Not run)

## Not run:
## Compare two aeids on same plots
tcplMakeAeidPlts(aeid = c(1,2), compare=T, lvl = 6)

## End(Not run)
```

tcplMakeChidMultiPlts *Create a .pdf with all dose-response plots for a given chid, 6 per page*

Description

tcplMakeChidMultiPlts Create a .pdf with all dose-response plots for a given chid

Usage

```
tcplMakeChidMultiPlts(
  chid,
  lvl = 4L,
  fname = NULL,
  odir = getwd(),
  clib = NULL,
  hitc.all = TRUE
)
```

Arguments

chid	Integer of length 1, the chemical id
lvl	Integer of length 1, the data level to use (4-7)
fname	Character, the filename
odir	The directory to save the .pdf file in
clib	Character, the chemical library to subset on, see tcplLoadClib for more information.
hitc.all	If FALSE, only plots with hitc==1 will be displayed

Details

tcplMakeChidMultiPlts provides a wrapper for [tcplMultiplot](#), allowing the user to produce PDFs with the curve plots without having to separately load all of the data and establish the PDF device.

If 'fname' is NULL, a default name is given by concatenating together assay information.

tcplMultiplot	<i>Plot summary fits based on fit and dose-response data</i>
---------------	--

Description

tcplMultiplot takes the dose-response and fit data and produces summary plot figures.

Usage

```
tcplMultiplot(dat, agg, flg = NULL, boot = NULL, browse = FALSE, hitc.all)
```

Arguments

dat	data.table, level 4 or level 5 data, see details.
agg	data.table, concentration-response aggregate data, see details.
flg	data.table, level 6 data, see details.
boot	data.table, level 7 data, see details.
browse	Logical, should browser() be called after every plot?
hitc.all	Logical, if FALSE, only plots with hitc==1 will be displayed

Details

The data for 'dat', 'agg', and 'flg' should be loaded using the [tcplLoadData](#) function with the appropriate 'lvl' parameter. See help page for tcplLoadData for more information.

If dat contains only one aeid, plots will be ordered by chemical name (chm). Otherwise, plots are ordered by assay endpoint name (aenm). ## While it is most likely the user will want to just save all of the plots ## to view in a PDF, the 'browse' parameter can be used to quickly view ## some plots.

tcplPlot	#----- <i>Plotting Function for tcpl</i>	<i>Generic</i>
----------	---	----------------

Description

tcplLoadData queries the tcpl databases and returns a plot for the given level and data type.

Usage

```
tcplPlot(
  lvl = 5,
  fld = "m4id",
  val = NULL,
  type = "mc",
  by = NULL,
  output = c("console", "pdf"),
  fileprefix = paste0("tcplPlot_", Sys.Date()),
  multi = FALSE,
  verbose = FALSE,
  nrow = NULL,
  ncol = NULL
)
```

Arguments

lvl	Integer of length 1, the level of data to load
fld	Character, the field(s) to query on
val	List, vectors of values for each field to query on. Must be in the same order as 'fld'.
type	Character of length 1, the data type, "sc" or "mc"
by	Parameter to divide files into e.g. aeid
output	how should the output be presented
fileprefix	prefix of filename
multi	Boolean, if multi is TRUE output 6 plots per page
verbose	By default FALSE, should a table with fitting parameters be included in the plot
nrow	Integer, number of rows in multiplot default of 2
ncol	Integer, number of columns in multiplot default of 3, 2 if verbose

Details

The data type can be either 'mc' for multiple concentration data, or 'sc' for single concentration data. Multiple concentration data will be loaded into the 'mc' tables, whereas the single concentration will be loaded into the 'sc' tables.

Setting 'lvl' to "agg" will return an aggregate table containing the m4id with the concentration-response data and m3id to map back to well-level information.

Leaving fld NULL will return all data.

Examples

```
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcpIConfList()
tcpIConfExample()

tcpIPlot(lvl = 4, fld = "m4id", val = c(18609966)) ## Create a level 4 plot

## Reset configuration
options(conf_store)
```

tcpIPlotFitc

Plot the fit category tree

Description

tcpIPlotFitc makes a plot showing the level 5 fit categories.

Usage

```
tcpIPlotFitc(fitc = NULL, main = NULL, fitc_sub = NULL)
```

Arguments

fitc	Integer, the fit categories
main	Character of length 1, the title (optional)
fitc_sub,	Integer, a subset of fit categories to plot

Note

Suggested device size (inches): width = 10, height = 7.5, pointsize = 9

Examples

```
## Not run:
## Plot visualization of fit categories for all level 5 data
tcplPlotFitc(fitc = tcplLoadData(5)$fitc)

## End(Not run)
```

tcplPlotFits

Plot summary fits based on fit and dose-response data

Description

tcplPlotFits takes the dose-response and fit data and produces summary plot figures.

Usage

```
tcplPlotFits(
  dat,
  agg,
  flg = NULL,
  boot = NULL,
  ordr.fitc = FALSE,
  browse = FALSE,
  cnst = NULL,
  orig.aeid = NULL,
  compare = F
)
```

Arguments

dat	data.table, level 4 or level 5 data, see details.
agg	data.table, concentration-response aggregate data, see details.
flg	data.table, level 6 data, see details.
boot	data.table, level 7 data, see details.
ordr.fitc	Logical, should the fits be ordered by fit category?
browse	Logical, should browser() be called after every plot?
cnst	Constant hline to draw on plot
orig.aeid	Original aeid list from tcplMakeAeidPlts to maintain order
compare	boolean to determine if aeids should be compared on same plot

Details

The data for 'dat', 'agg', and 'flg' should be loaded using the `tcplLoadData` function with the appropriate 'lvl' parameter. See help page for `tcplLoadData` for more information.

Supplying level 4 data for the 'dat' parameter will result in level 4 plots. Similarly, supp

If fits are not ordered by fit category, they will be ordered by chemical ID. Inputs with multiple assay endpoints will first be ordered by assay endpoint ID.

Examples

```
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcplConfList()
tcplConfDefault()

## tcplPlotFits needs data.tables supplying the concentration/response
## data stored in mc4_agg, as well as the fit information from mc4 or mc5.
## Additionally, tcplPlotFits can take level 6 data from mc6 and add the
## flag information to the plots. The following shows how to make level 5
## plots. Adding the 'flg' parameter would result in level 6 plots, and
## loading level 4, rather than level 5 data, would result in level 4 plots.

l5 <- tcplLoadData(lvl = 5, fld = "m4id", val = 18609966)
l4_agg <- tcplLoadData(lvl = "agg", fld = "m4id", val = 18609966)

## Not run:
pdf(file = "tcplPlotFits.pdf", height = 6, width = 10, pointsize = 10)
tcplPlotFits(dat = l5, agg = l4_agg)
graphics.off()

## End(Not run)

## While it is most likely the user will want to just save all of the plots
## to view in a PDF, the 'browse' parameter can be used to quickly view
## some plots.

## Start by identifying some sample IDs to plot, then call tcplPlotFits with
## a subset of the data. This browse function is admittedly clunky.
bpa <- tcplLoadChem(field = "chnm", val = "Bisphenol A")[ , spid]
l5_sub <- l5[spid %in% bpa]
## Not run:
tcplPlotFits(dat = l5_sub,
             agg = l4_agg[m4id %in% l5_sub$m4id],
             browse = TRUE)

## End(Not run)

## Reset configuration
options(conf_store)
```

tcplPlotlyPlot	<i>tcplPlotlyPlot</i>
----------------	-----------------------

Description

tcplPlotlyPlot

Usage

```
tcplPlotlyPlot(dat, lvl = 5)
```

Arguments

dat	data table with all required conc/resp data
lvl	integer level of data that should be plotted level 4 - all fit models level 5 - all fit models and winning model with hitcall level 6 - include all flags

Value

A plotly plot

tcplPlotM4ID	<i>Plot fit summary plot by m4id</i>
--------------	--------------------------------------

Description

tcplPlotM4ID creates a summary plots for the given m4id(s) by loading the appropriate data from the tcpl databases and sending it to [tcplPlotFits](#)

Usage

```
tcplPlotM4ID(m4id, lvl = 4L)
```

Arguments

m4id	Integer, m4id(s) to plot
lvl	Integer, the level of data to plot

Details

A level 4 plot (`lvl = 4`) will plot the concentration series and the applicable curves, without an indication of the activity call or the winning model. Level 4 plots can be created without having done subsequent processing.

Level 5 plots include the level 4 information with the activity call and model selection. The winning model will be highlighted red in the side panel containing the summary statistics. Level 6 plots, in addition to all of the level 4 and 5 information, include the positive flag IDs. If the flag has an associated value, the value will be in parentheses following the flag ID. Level 7 plots in addition to all of the level 4, 5, and 6 information, include the AC50 confidence interval and hit percentage information from bootstrapping.

See Also

[tcplPlotFits](#), [tcplMakeAeidPlts](#)

Examples

```
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcplConfList()
tcplConfExample()

tcplPlotM4ID(m4id = 18609966, lvl = 4) ## Create a level 4 plot
tcplPlotM4ID(m4id = 18609966, lvl = 5) ## Create a level 5 plot
tcplPlotM4ID(m4id = 18609966, lvl = 6) ## Create a level 6 plot

#' ## Reset configuration
options(conf_store)
```

tcplPlotPlate

Plot plate heatmap

Description

tcplPlotPlate generates a heatmap of assay plate data

Usage

```
tcplPlotPlate(dat, apid, id = NULL, quant = c(0.001, 0.999))
```

Arguments

dat	data.table containing tcpl data
apid	Character of length 1, the apid to plot
id	Integer of length 1, the assay component id (acid) or assay endpoint id (aeid), depending on level. Only need to specify for multiplexed assays when more than one acid/aeid share an apid.
quant	Numeric vector, the range of data to include in the legend

Details

The legend represents the range of the data supplied to `dat`, for the applicable ID. The additional horizontal lines on the legend indicate the range of the plotted plate, to show the relation of the plate to the assay as a whole. A plot with a legend specific for the given apid can be created by only supplying the data for the apid of interest to `'dat'`.

The `quant` parameter, by default including 99.8 allows for extreme outliers without losing resolution. Outliers in either direction will be highlighted with a dark ring, as seen in the example. A NULL value for `'quant'` will not restrict the data at all, and will use the full range for the legend.

Wells with a well quality of 0 (only applicable for level 1 plots), will have an "X" through their center.

Note

For the optimal output size, use `width = 10, height = 10*(2/3), pointsize = 10, units = "in"`

Examples

```
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcplConfList()
tcplConfDefault()

d1 <- tcplLoadData(lvl = 1, fld = "acid", val = 1)
## Not run:
tcplPlotPlate(dat = d1, apid = "09Apr2014.Plate.17")

## End(Not run)

## Reset configuration
options(conf_store)
```

 tcplPrep0tpt

Map assay/chemical ID values to annotation information

Description

`tcplPrep0tpt` queries the chemical and assay information from the `tcpl` database, and maps the annotation information to the given data.

Usage

```
tcplPrep0tpt(dat, ids = NULL)
```

Arguments

<code>dat</code>	data.table, output from <code>tcplLoadData</code>
<code>ids</code>	Character, (optional) a subset of ID fields to map

Details

tcp1Prep0tpt is used to map chemical and assay identifiers to their respective names and annotation information to create a human-readable table that is more suitable for an export/output.

By default the function will map sample ID (spid), assay component id (acid), and assay endpoint ID (aeid) values. However, if 'ids' is not null, the function will only attempt to map the ID fields given by 'ids.'

Value

The given data.table with chemical and assay information mapped

Examples

```
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcp1ConfList()
tcp1ConfExample()

## Load some example data
d1 <- tcp1LoadData(1)

## Check for chemical name in 'dat'
"chnm" %in% names(d1) ## FALSE

#' ## Map all annotations
d2 <- tcp1Prep0tpt(d1) ##
"chnm" %in% names(d2) ## TRUE
"acnm" %in% names(d2) ## TRUE

## Map chemical annotation only
d3 <- tcp1Prep0tpt(d1, ids = "spid")
"chnm" %in% names(d3) ## TRUE
"acnm" %in% names(d3) ## FALSE

## Reset configuration
options(conf_store)
```

tcp1Run

Perform data processing

Description

tcp1Run is the function for performing the data processing, for both single-concentration and multiple-concentration formats.

Usage

```
tcplRun(  
  asid = NULL,  
  slvl,  
  elvl,  
  id = NULL,  
  type = "mc",  
  mc.cores = NULL,  
  outfile = NULL,  
  runname = NULL  
)
```

Arguments

asid	Integer, assay source id
slvl	Integer of length 1, the starting level to process
elvl	Integer of length 1, the ending level to process
id	Integer, rather than assay source id, the specific assay component or assay end-point id(s) (optional)
type	Character of length 1, the data type, "sc" or "mc"
mc.cores	Integer of length 1, the number of cores to use, set to 1 when using Windows operating system
outfile	Character of length 1, the name of the log file (optional)
runname	Character of length 1, the name of the run to be used in the outfile (optional)

Details

The `tcplRun` function is the core processing function within the package. The function acts as a wrapper for individual processing functions, (ie. `mc1`, `sc1`, etc.) that are not exported. If possible, the processing is done in parallel by 'id' by utilizing the `mclapply` function within the parallel package.

If `slvl` is less than 4, 'id' is interpreted as `acid` and if `slvl` is 4 or greater 'id' is interpreted as `aeid`. Must give either 'asid' or 'id'. If an id fails no results get loaded into the database, and the id does not get placed into the cue for subsequent level processing.

The 'type' parameter specifies what type of processing to complete: "mc" for multiple-concentration processing, and "sc" for single-concentration processing.

Value

A list containing the results from each level of processing. Each level processed will return a named logical vector, indicating the success of the processing for the id.

tcplSubsetChid	<i>Subset level 5 data to a single sample per chemical</i>
----------------	--

Description

tcplSubsetChid subsets level 5 data to a single tested sample per chemical. In other words, if a chemical is tested more than once (a chid has more than one spid) for a given assay endpoint, the function uses a series of logic to select a single "representative" sample.

Usage

```
tcplSubsetChid(dat, flag = TRUE, type = "mc")
```

Arguments

dat	data.table, a data.table with level 5 data
flag	Integer, the mc6_mthd_id values to go into the flag count, see details for more information
type	Character of length 1, the data type, "sc" or "mc"

Details

tcplSubsetChid is intended to work with level 5 data that has chemical and assay information mapped with [tcplPrep0tpt](#).

To select a single sample, first a "consensus hit-call" is made by majority rule, with ties defaulting to active. After the chemical-wise hit call is made, the samples corresponding to to chemical-wise hit call are logically ordered using the fit category, the number of the flags, and the modl_ga, then the first sample for every chemical is selected.

The flag param can be used to specify a subset of flags to be used in the flag count. Leaving flag TRUE utilize all the available flags. Setting flag to FALSE will do the subsetting without considering any flags.

Value

A data.table with a single sample for every given chemical-assay pair.

See Also

[tcplPrep0tpt](#)

Examples

```
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcplConfList()
tcplConfExample()
```

```
## Load the example level 5 data
d1 <- tcplLoadData(lvl = 5, fld = "aeid", val = 797)
d1 <- tcplPrepOtppt(d1)

## Subset to an example of a duplicated chid
d2 <- d1[chid == 20182]
d2[ , list(m4id, hitc, fitc, modl_ga)]

## Here the consensus hit-call is 1 (active), and the fit categories are
## all equal. Therefore, if the flags are ignored, the selected sample will
## be the sample with the lowest modl_ga.
tcplSubsetChid(dat = d2, flag = FALSE)[ , list(m4id, modl_ga)]

## Reset configuration
options(conf_store)
```

tcplVarMat

Create chemical by assay matrices

Description

tcplVarMat creates chemical by assay matrices.

Usage

```
tcplVarMat(
  chid = NULL,
  aeid = NULL,
  add.vars = NULL,
  row.id = "code",
  flag = TRUE,
  cyto.pars = list(),
  include.na.chid = FALSE,
  odir = NULL,
  file.prefix = NULL
)
```

Arguments

chid	Integer, chemical ID values to subset on
aeid	Integer, assay endpoint ID values to subset on
add.vars	Character, mc4 or mc5 field(s) not included in the standard list to add additional matrices
row.id	Character, the chemical identifier to use in the output
flag	Integer or Logical of length 1, passed to tcplSubsetChid
cyto.pars	List, named list of arguments passed to tcplCytoPt

<code>include.na.chid</code>	Logical of length 1, whether to include the chemicals not listed in the tcp1 databases (ie. controls)
<code>odir</code>	Directory to write comma separated file(s)
<code>file.prefix</code>	Character of length 1, prefix to the file name when <code>odir</code> is not NULL

Details

The `tcp1VarMat` function is used to create chemical by assay matrices for different parameters. The standard list of matrices returned includes:

1. "modl_ga" – The logAC50 (in the gain direction) for the winning model.
2. "hitc" – The hit-call for the winning model.
3. "m4id" – The m4id, listing the concentration series selected by `tcp1SubsetChid`.
4. "zscore" – The z-score based on the output from `tcp1CytoPt`. The formula used for calculating the z-score is $-(modl_ga - cyto_pt)/global_mad$
5. "tested" – 1 or 0, 1 indicating the chemical/assay pair was tested in either the single- or multiple-concentration format
6. "tested_sc" – 1 or 0, 1 indicating the chemical/assay pair was tested in the single-concentration format
7. "tested_mc" – 1 or 0, 1 indicating the chemical/assay pair was tested in the multiple-concentration format
8. "ac50" – a modified AC50 table (in non-log units) where assay/chemical pairs that were not tested, or tested and had a hitcall of 0 or -1 have the value $1e6$.
9. "neglogac50" – $-\log(AC50/1e6)$ where assay/chemical pairs that were not tested, or tested and had a hitcall of 0 or -1 have the value 0.

To add additional matrices, the `'add.vars'` parameter can be used to specify the fields from the `mc4` or `mc5` tables to create matrices for.

When more than one sample is included for a chemical/assay pair, `tcp1VarMat` aggregates multiple samples to a chemical level call utilizing `tcp1SubsetChid`.

By setting `odir` the function will write out a csv with, naming the file with the convention: `"var_Matrix_date.csv"` where `'var'` is the name of the matrix. A prefix can be added to the output files using the `'file.prefix'` parameter.

When a concentration series has a sample id not listed in the tcp1 database, and `'include.na.chid'` is TRUE, the rowname for that series will be the concatenation of "SPID_" and the `spid`. Note, if the user gives a subset of `chid` values to the `'chid'` parameter, `'include.na.chid'` will be set to FALSE with a warning.

The `tcp1VarMat` function calls both `tcp1SubsetChid` and `tcp1CytoPt` (which separately calls `tcp1SubsetChid`). The input for the `tcp1VarMat` `'flag'` parameter is passed to the `tcp1SubsetChid` call used to parse down the data to create the matrices. The `tcp1SubsetChid` called within `tcp1CytoPt` (to parse down the cytotoxicity data used to define the "zscore" matrix) can be modified by passing a separate `'flag'` element in the list defined by the `'cyto.pars'` parameter.

Value

A list of chemical by assay matrices where the rownames are given by the 'row.id' parameter, and the colnames are given by assay endpoint name (aenm).

See Also

[tcplSubsetChid](#)

Examples

```
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcplConfList()
TCPLlite <- file.path(system.file(package = "tcpl"), "example")
tcplConf(db = TCPLlite, user = NA, host = NA, drvr = "tcplLite")
## Not run:
## Demonstrate the returned values. Note with no "burst" assays defined in
## the example database, the user must provide which aeid values to use
## in calculating the cytotoxicity distributions for the 'zscore' matrix.
tcplVarMat(chid = 1:5, cyto.pars = list(aeid = 1:2))

## Other changes can be made
tcplVarMat(chid = 1:5, row.id = "chnm", cyto.pars = list(aeid = 1:2))
tcplVarMat(chid = 1:5, add.vars = "max_med", cyto.pars = list(aeid = 1:2))

## End(Not run)
## Reset configuration
options(conf_store)
```

tcplWriteData

Write screening data into the tcpl databases

Description

tcplWriteData takes a data.table with screening data and writes the data into the given level table in the tcpl databases.

Usage

```
tcplWriteData(dat, lvl, type)
```

Arguments

dat	data.table, the screening data to load
lvl	Integer of length 1, the data processing level
type	Character of length 1, the data type, "sc" or "mc"

Details

This function appends data onto the existing table. It also deletes all the data for any acids or acids dat contains from the given and all downstream tables.

The data type can be either 'mc' for multiple concentration data, or 'sc' for single concentration data. Multiple concentration data will be loaded into the level tables, whereas the single concentration will be loaded into the single tables.

Note

This function is not exported and is not intended to be used by the user. The user should only write level 0 data, which is written with [tcplWriteLv10](#).

See Also

[tcplCascade](#), [tcplAppend](#), [tcplWriteLv10](#)

tcplWriteLv10

Write level 0 screening data into the tcpl databases

Description

tcplWriteLv10 takes a data.table with level 0 screening data and writes the data into the level 0 tables in the tcpl databases.

Usage

```
tcplWriteLv10(dat, type)
```

Arguments

dat	data.table, the screening data to load
type	Character of length 1, the data type, "sc" or "mc"

Details

This function appends data onto the existing table. It also deletes all the data for any acids or acids dat contains from the given and all downstream tables.

Before writing any data the function maps the assay component source name(s) (acsn) to assay component id (acid), ensures the proper class on each field and checks for every test compound sample id (spid where wllt == "t") in the tcpl chemical database. If field types get changed a warning is given listing the affected fields and they type they were coerced to. If the acsn(s) or spid(s) do not map to the tcpl databases the function will return an error and the data will not be written.

The data type can be either 'mc' for multiple concentration data, or 'sc' for single concentration data. Multiple concentration data will be loaded into the level tables, whereas the single concentration will be loaded into the single tables.

Note

This function should only be used to load level 0 data.

See Also

[tcplCascade](#), [tcplAppend](#)

write_lvl_4

Write level 4 with updated schema

Description

Write level 4 with updated schema

Usage

```
write_lvl_4(dat)
```

Arguments

dat output of tcplfit2 that has been unnested into a data.table

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